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(54) Title: LUNG CANCER MARKER

(57) Abstract

The present invention discloses an isolated and purified nucleic acid sequence and corresponding amino acid sequence to a novel protein specific for human lung cancer cells. This gene is expressed at a much higher level in these cells than in normal lung cells, other normal tissues and other tumor cell lines tested. Also disclosed are three additional recombinant forms of this gene and protein, in the first two cases a membrane spanning region is removed and in the third case an amino acid is changed by in vitro mutagenesis. Also disclosed are nucleic acid probes for the detection of lung cancer cells from tissue biopsy and body fluids such as serum, sputum and bronchial washings. A method for expressing the antigen in a host cell and its subsequent use as an immunogen in antibody production for test applications is described. An ELISA test to measure shed antigen present in patient samples as well as an enzyme test to measure activity in specimens also is described.

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LUNG CANCER MARKER

TECHNICAL FIELD

The invention relates to genes and proteins specific for certain cancers and methods for their detection.

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BACKGROUND OF THE INVENTION

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Lung cancer is the most common form of cancer in the world. Estimates for the year 1985 indicate that there were about 900,000 cases of lung cancer worldwide. (Parkin, et al., "Estimates of the worldwide incidence of eighteen major cancers in 1985," Int J Cancer 1993; 54:594-606). For the United States alone, 1993 projections placed the number of new lung cancer cases at 170,000, with a mortality of about 88%. (Boring, et al., "Cancer statistics," CA Cancer J Clin 1993; 43:7-26). Although the occurrence of breast cancer is slightly more common in the United States, lung cancer is second behind prostate cancer for males and third behind breast and colorectal cancers for women. Yet, lung cancer is the most common cause of cancer deaths.

The World Health Organization classifies lung cancer into four major histological types: (1) squamous cell carcinoma (SCC), (2) adenocarcinoma, (3) large cell carcinoma, and (4) small cell lung carcinoma (SCLC). (The World Health Organization, "The World Health Organization histological typing of lung tumours," Am J Clin Pathol 1982; 77:123-136). However, there is a great deal of tumor heterogeneity even within the various subtypes, and it is not uncommon for lung cancer to have features of more than one morphologic subtype. The term non-small cell lung carcinoma (NSCLC) includes squamous, adenocarcinoma and large cell carcinomas.

Typically, a combination of X-ray and sputum cytology is used to diagnose lung cancer. Unfortunately, by the time a patient seeks medical help for their symptoms, the cancer is at such an advanced state it is usually incurable. Cancer Facts and Figures (based on rates from

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NCI SEER Program 1977-1981), New York: American Cancer Society, 1986). Routine large-scale radiologic or cytologic screening of smokers has been investigated. Studies concluded that cytomorphological screening did not significantly reduce the mortality rate from lung cancer and was not recommended for routine use. ("Early lung cancer detection: summary & conclusions," Am Rev Respir Dis 1984; 130:565-70). However, in a subpopulation of patients where the cancer is diagnosed at a very early stage and the lung is surgically resectioned, there is a 5-year survival rate of 70-90%. (Flehinger, et al., "The effect of surgical treatment on survival from early lung cancer," Chest; 1992, 101:1013-1018; Melamed, et al., "Screening for early lung cancer: results of the Memorial Sloan-Kettering Study in New York, " Chest; 1984 86:44-53). Therefore, research has focused on early detection of tumor markers before the cancer becomes clinically apparent and while the cancer is still localized and amenable to therapy.

The identification of antigens associated with lung cancer has stimulated considerable interest because of their use in screening, diagnosis, clinical management, and potential treatment of lung cancer. International workshops have attempted to classify the lung cancer antigens into 15 possible clusters that may define histologic origins. (Souhami, et al., "Antigens of lung cancer: results of the second international workshop on lung cancer antigens," JNCI 1991; 83:609-612). As of 1988, more than 200 monoclonal antibodies (MAb) have been reported to react with human lung tumors. (Radosevich, et al., "Monoclonal antibody assays for lung cancer," In:

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Cancer Diagnosis in Vitro Using Monoclonal Antibodies.
Edited by H. A. Kupchik. New York: Marcel Dekker, 1988).

MAbs for lung cancer were first developed to distinguish NSCLC from SCLC. (Mulshine, et al., "Monoclonal antibodies that distinguish nonsmall-cell from small-cell lung cancer," *J Immunol* 1983; 121:497-502). In most cases, the identity of the cell surface antigen with which a particular antibody reacts is not known, or has not been well characterized. (Scott, et al., "Early lung cancer detection using monoclonal antibodies," In: *Lung Cancer*. Edited by J.A. Roth, J.D. Cox, and W.K. Hong. Boston: Blackwell Scientific Publications, 1993).

MAbs have been used in the immunocytochemical staining of sputum samples to predict the progression of lung cancer. (Tockman, et al., "Sensitive and specific monoclonal antibody recognition of human lung cancer antigen on preserved sputum cells: a new approach to early lung cancer detection, " J Clin Oncol 1988; 6:1685-1693). In the study, two MAbs were utilized, 624H12 which binds a glycolipid antigen expressed in SCLC and 703D4 which is directed to a protein antigen of NSCLC. Of the sputum specimens from participants who progressed to lung cancer, two-thirds showed positive reactivity with either the SCLC or the NSCLC MAb. In contrast, of those that did not progress to lung cancer, 35 of 40 did not react with the SCLC or NSCLC Mab. This study suggests the need for the development of additional early detection targets to discover the onset of malignancy at the earliest possible stage.

Carcinoembryonic antigen (CEA) is a frequently studied tumor marker of cancer including lung cancer.

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(Nutini, et al., "Serum NSE, CEA, CT, CA 15-3 levels in human lung cancer," Int J Biol Markers 1990; 5:198-202). Squamous cell carcinoma antigen is another established serum marker. (Margolis, et al., "Serum tumor markers in non-small cell lung cancer," Cancer 1994; 73:605-609.). Other serum antigens for lung cancer include antigens recognized by MAbs 5E8, 5C7, and 1F10, the combination of which distinguishes between patients with lung cancer from those without. (Schepart, et al., "Monoclonal antibodymediated detection of lung cancer antigens in serum, " Am Rev Respir Dis 1988; 138:1434-8) Furthermore, the combination of 5E8, 5C7 and 1F10 was more sensitive, specific and accurate for identifying NSCLC when compared to results from a combination of the CEA and squamous cell carcinoma antigen tests. (Margolis, et al., Cancer 1994; 73:605-609).

Serum CA 125, initially described as an ovarian cancer-associated antigen, has been investigated for its use as a prognostic factor in NSCLC. (Diez, et al., "Prognostic significance of serum CA 125 antigen assay in patients with non-small cell lung cancer," Cancer 1994; 73:136876). The study determined that the preoperative serum level of CA 125 antigen is inversely correlated with survival and tumor relapse in NSCLC.

Despite the numerous examples of MAb applications, none has yet emerged that has changed clinical practice.

(Mulshine, et al., "Applications of monoclonal antibodies in the treatment of solid tumors," In: Biologic Therapy of Cancer. Edited by V.T. Devita, S. Hellman, and S.A.

Rosenberg. Philadelphia: JB Lippincott, 1991, pp. 563-588). MAbs alone may not be the answer to early detection

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because there has only been moderate success with immunologic reagents for paraffin-embedded tissue. Secondly, lung cancer may express features that cannot be differentiated by antibodies; for example, chromosomal deletions, gene amplification, or translocation and alteration in enzymatic activity.

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After the gene to the MAb recognized surface antigen has been cloned, cytogenetic and molecular techniques may provide powerful tools for screening, diagnosis, management and ultimately treatment of lung cancer. An example of a lung cancer antigen that has been cloned is the adenocarcinoma-associated antigen. This antigen, recognized by KS1/4 MAb, is an epithelial malignancy/epithelial tissue glycoprotein from the human lung adenocarcinoma cell line UCLA-P3. (Strand, et al., "Molecular cloning and characterization of a human adenocarcinoma/epithelial cell surface antigen complementary DNA," Cancer Res 1989; 49:314-317). antigen has been found on all adenocarcinoma cells tested and in various corresponding normal epithelial cells. Northern blot analysis indicated that transcription of the adenocarcinoma-associated antigen was detected in RNA isolated from normal colon but not in RNA isolated from normal lung, prostate, or liver. Therefore identification of adenocarcinoma-associated antigen in lung cells may prove to be diagnostic for adenocarcinoma.

The cloning of CEA and the nonspecific crossreacting antigen (NCA) has allowed the development of specific DNA probes which discriminate their expression in lung cancer at the mRNA level. (Hasegawa, et al., "Nonspecific crossreacting antigen (NCA) is a major member of the CEA-related gene family expressed in lung cancer," Br J Cancer

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1993; 67:58-65). NCA is a component of the CEA gene family in lung cancer and is also recognized by anti-CEA antibodies, especially polyclonal antibodies. Because of the crossreactivity, investigations to analyze CEA and NCA separately in lung disease had been difficult. The use of DNA probes determined that lung cancer cells fall into three different types according to their CEA and/or NCA expression by Northern blot analysis. Specifically, lung cancers expressed both CEA and NCA mRNA, only NCA mRNA, or neither mRNA. CEA-related mRNA expression was always accompanied by NCA mRNA expression and there were no cases of CEA mRNA expression alone. The separate assessment of CEA and NCA expression in lung cancers may be important in determining the prognosis of lung cancers because the antigens have been described as cell-cell adhesion molecules and may play a role in cancer metastasis.

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Another method to detect the presence of an antigen gene or its mRNA in specific cells or to localize an antigen gene to a specific locus on a chromosome is in situ hybridization. In situ hybridization uses nucleic acid probes that recognize either repetitive sequences on a chromosome or sequences along the whole chromosome length or chromosome segments. By tagging the probes with radioisotopes or color detection systems, chromosome regions can be identified within the cell. Investigations using in situ hybridization have demonstrated numerical chromosomal abnormalities in samples from human tumors, including bladder, neuroectodermal, breast, gastric and lung cancer tumors. (Kim, et al., "Interphase cytogenetics in paraffin sections of lung tumors by non-isotopic in situ hybridization. Mapping Genotype/phenotype heterogeneity," Am J Pathol 1993; 142:307-317).

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Fluorescence in situ hybridization (FISH) allows cells to be stained so that genetic aberrations resulting in changes in gene copy number or structure can be quantitated by fluorescence microscopy. In this technique, a chemically labeled single-stranded nucleic acid probe homologous to the target nucleic acid sequence is annealed to denatured nucleic acid contained in target The cells may be mounted on a microscope slide, in suspension or prepared from paraffin-embedded material. Treating the chemically modified probes with a fluorescent ligand makes the bound probe visible. FISH has been used for (1) detection of changes in gene copy number and gene structure; (2) detection of genetic changes, even in low frequency subpopulations; and (3) detection and measurement of the frequency of residual malignant cells. (Gray, et al., "Molecular cytogenetics in human cancer diagnosis," Cancer 1992; 69:1536-1542).

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Other molecular markers for lung cancer include oncogenes and tumor suppressor genes. Dominant oncogenes are activated by mutation and lead to deregulated cellular growth. Such genes code for proteins that function as growth factors, growth factor receptors, signal transducing proteins and nuclear proteins involved in transcriptional regulation. Amplification, mutation, and translocations have been documented in many different cancer cells and have been shown to lead to gene activation or overexpression.

The ras family of oncogenes comprises a group of membrane associated GTP-binding proteins thought to be involved in signal transduction. Mutations within the ras oncogenes, resulting in sustained growth stimulation, have been identified in 15 to 30% of human NSCLC. (Birrer, et

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al., "Application of molecular genetics to the early diagnosis and screening of lung cancer," Cancer 1992; 52suppl; 2658s-2664s). Patients with tumors containing ras mutations had decreased survival compared with patients whose tumors had no ras mutations. Polymerase chain reaction (PCR) amplification of ras genes can be analyzed to determine the presence of mutations by several methods: (a) differential hybridization of ³²P-labeled mutated oligonucleotides; (b) identification of new restriction enzyme sites created by the activating mutation; (c) single-strand conformational polymorphisms; and (d) nucleic acid sequencing. These methods combined with PCR technology could allow detection of an activated ras gene from sputum specimens.

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Another family of dominant oncogenes, the erb B family, has been found to be abnormally expressed in lung cancer cells. This group codes for membrane-associated tyrosine kinase proteins and contains erb B1, the gene coding for the epidermal growth factor (EGF) receptor, and erb B2 (also called Her-2/neu). The erb B1 gene has been found to be amplified in NSCLC (up to 20% of squamous cell tumors), while the EGF receptor has been shown to be overexpressed in many NSCLC cells (approximately 90% of squamous cell tumors, 20 to 75% of adenocarcinomas, and rarely in large cell or undifferentiated tumors). (Birrer, et al., Cancer 1992: 52 suppl; 2658s-2664s). Amplification of the related oncogene erb B2 (Her-2/neu) occurs infrequently in lung cancer but is a negative prognostic factor in breast cancer. However, overexpression of the erb B2 protein product, p185neu, occurs in some NSCLC and may be related to poor prognosis. (Kern, et al., "p185" expression in human lung

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adenocarcinomas predicts shortened survival," *Cancer Res* 1990; 50:5184-5191).

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A third family of dominant oncogenes involved in lung cancer is the myc family. These genes encode nuclear phosphoproteins, which have potent effects on cell growth and which function as transcriptional regulators. Unlike ras genes, which are activated by point mutations in lung cancer cells, the myc genes are activated by overexpression of the cellular myc genes, either by gene amplification or by rearrangements, each ultimately leading to increased levels of myc protein. Amplification of the normal myc genes is seen frequently in SCLC and rarely in NSCLC.

The loss or inactivation of tumor suppressor genes may also be important steps in the pathway leading to invasive cancer. Tumor suppressor genes function normally to suppress cellular proliferation, and since they are recessive oncogenes, mutations or deletions must occur in both alleles of these genes before transformation occurs.

A phosphoprotein p53, which is encoded by a gene located on chromosome 17p, suppresses transformation in its wild-type state. While in its mutant state, p53 acts as a dominant oncogene. p53 functions in DNA binding and transcription activation. Mutations of p53 have been found in many human cancers including colon, breast, brain and lung cancer cells. (Birrer, et al., Cancer Res.(suppl) 1992, 52:2658s-2664s). In NSCLC cell lines, p53 mutations have been found at a rate of up to 74%. (Mitsudomi, et al., "p53 gene mutations in non-small-cell lung cancer cell lines and their correlation with the

presence of ras mutations and clinical features," Oncogene 1992; 7:171-180).

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Despite all of the advances made in the area of lung cancer, medical and surgical intervention has resulted in little change in the 5-year survival rate for lung cancer patients. Early detection holds the greatest hope for successful intervention. There remains a need for a practical method to diagnose lung cancer as close to its inception as possible. In order for early detection to be feasible, it is important that specific markers be found and their sequences elucidated.

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A lung cancer marker antigen, specific for NSCLC, has now been found, sequenced, and cloned. The antigen is useful in methods for detection of non-small cell lung cancer and for potential production of antibodies and probes for treatment compositions.

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BRIEF DESCRIPTION OF THE DRAWING

FIGURE 1 depicts the alignment of the amino acid sequence of HCAVIII with previously described carbonic anhydrases. Conserved amino acids are shown in bold.

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SUMMARY OF THE INVENTION

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The invention concerns a lung cancer antigen (HCAVIII) gene specific for non-small cell lung cancer.

In one embodiment, the invention relates to a substantially purified nucleic acid (SEQ ID NO:1) encoding the pre-protein sequence shown in SEQ ID NO:2.

In other embodiments, the invention relates to cDNAs which encode the mature form of the protein (SEQ ID NO:4), or a truncated form of the protein lacking the transmembrane domain (SEQ ID NO:13 and SEQ ID NO:15), or a protein in which one or more of the amino acids in the phosphorylation region have been altered to affect that function, an example of which is shown in SEQ ID NO:18.

In other embodiments, proteins encoded by the cDNA of SEQ ID NO:1, SEQ ID NO:3, SEQ ID NO:5, SEQ ID NO:12, SEQ ID NO:14, and SEQ ID NO:17 are provided.

In another aspect, the invention relates to a recombinant DNA clone for HCAVIII.

In further aspects of the invention, expression vectors for HCAVIII and modifications thereof are an object.

The invention further relates to methods of detecting lung cancer.

In one aspect an *in situ* hybridization technique is provided. In another aspect, a fluorescence *in situ* hybridization technique is provided. In a further aspect, an ELISA assay is provided. In another aspect, detection of carbonic anhydrase activity which correlates with lung cancer antigen is provided.

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DETAILED DESCRIPTION OF THE INVENTION

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The nucleic acid sequence coding for a cell surface protein (said protein hereinafter designated HCAVIII) which is highly specific for non-small cell lung cancer cells has now been obtained. This gene sequence will facilitate detection and treatment of the disease, which to date has often proven difficult.

The HCAVIII cDNA in the vector pLC56 has been sequenced and characterized including the entire coding region and substantially all of the upstream and downstream non-translated regions. The cDNA in pLC56 was sequenced on both strands from exonuclease III-generated deletions and subsequent subcloning into M13 vectors or directly from the cloning vectors using the di-deoxy method and a SEQUENASE ® Version 2.0 kit (U.S. Biochemicals, Cleveland, OH). Additional regions of DNA were subcloned as small restriction fragments into the same vectors for sequence analysis. Overlapping segments were ordered using MacVector Align software (Kodak/IBI Technologies, New Haven CT). SEQ ID NO:1 represents the cDNA encoding HCAVIII and a presumed signal peptide. ID NO:2 represents the signal peptide (amino acid residues -29 to -1) followed by the mature protein (amino acid residues 1 to 325). As predicted from the cDNA sequence in pLC56, a protein of about 354 amino acids is encoded with the predictive size of 39448 daltons. A hydrophilicity plot (MacVector software, Kodak/IBI Technologies) of this protein provided strong evidence of a leader peptide at the N-terminus and a membrane-spanning segment near the C-terminus. The membrane-spanning segment provides evidence that this protein is membrane bound, as also predicted by its positive selection with

panning methodology (See Watson, et al., Recombinant DNA, 2nd ed., pp. 115-116, 1992). The cleavage site of the signal as predicted by von Heijne (von Heijne, Gunnar, Nucleic Acids Res 1986; 14:4683-4690) is 29 amino acids down from the N-terminus methionine. SEQ ID NO:3 corresponds approximately to the coding region of the mature polypeptide. The subsequent "mature" protein is proposed to be 325 amino acids, initiating with serine, and of a calculated 36401 daltons and a pI of 6.42 (SEQ ID NO:4).

Homology searches against NCBI BlastN or BlastX version 1.3.12MP (National Center for Biotechnology Information, Bethesda, MD) provided evidence the gene and protein are novel, not previously identified in either database. (Altschul, et al., "Basic local alignment search tool," J Mol Biol 1990; 215:403-410). Additional searches against another database (Entrez, version 9) gave similar results.

The isolation of a second cDNA encoding HCAVIII permitted the identification of new sequences within the 5'-and 3'-prime untranslated regions of this gene. SEQ ID NO:5, a cDNA encoding HCAVIII and a portion of the 5' and 3' nontranslated regions, has substantial identity with SEQ ID NO:1 (positions 1-1104 of SEQ ID NO:1 are identical to positions 85-1188 of SEQ ID NO:5). The encoded protein is listed in SEQ ID NO:6 and is identical with SEQ ID NO:2. Homology searches of NCBI BlastN against SEQ ID NO:5 showed these gene sequences have not been previously identified. SEQ ID NO:7 represents additional cDNA sequences of the 3' nontranslated region of the HCAVIII gene located downstream from the sequences depicted in SEQ ID NO:5. Homology searches against the same data base

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identified two clones with homology to SEQ ID NO:7. Both sequences are expressed sequence tags (EST), the first EST04899 (345 bp) and the second HUMGS04024 (466 bp).

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Alignment searches indicate this protein shares common features with the seven human carbonic anhydrase proteins previously identified. However, as described below, certain structural features distinct to HCAVIII exist that may confer unique properties to this protein and a role in the transformation pathway to tumorgenicity. This group of enzymes catalyze the hydration of carbon dioxide

 $CO_2 + H_2O = HCO_3 + H^*$

and in reverse the dehydration of HCO3-. This protein is identified as a carbonic anhydrase (CA) based on the conservation of amino acids at positions critical for the binding of Zn^{+2} , and the catalysis of CO_2 , as well as numerous other conserved amino acids (see Fig. 1). The protein is 34 to 64 amino acids longer (at the C-terminus) than any previously reported carbonic anhydrase by virtue of the membrane-spanning region also found in HCAIV and an additional approximate 30 amino acids contained in the cytoplasmic side of the cell and apparently missing in other human CA isoforms. In addition, this intracellular domain contains a phosphorylation site recognized by protein kinase C and other kinases, as defined by the motif "Arg-Arg-Lys-Ser" (SEQ ID NO:8 and SEQ ID NO:9) (amino acid residues 1-4 in SEQ ID NO:9 and amino acid residues 299-302 in SEQ ID NO:2, SEQ ID NO:4 and SEQ ID NO:6).

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Interestingly, this motif is found only in HCAVIII, and at a functionally significant site, i.e., within the cytosol. A surface cleft essential for enzymatic function present on other carbonic anhydrases is conserved for this protein, suggesting that this protein will also confer enzymatic activity. Five possible N-glycosylation sites are predicted by the primary amino acid sequence and the motif "Asn-Xaa-Ser (Thr)", beginning at amino acid residues -2, 51, 133, 151, and 202 in SEQ ID NO:2, respectively.

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HCAVIII is expressed at a much higher level in a nonsmall cell lung cancer cell line (A549) than in normal lung tissue, other normal tissues, and other tumor cell lines which makes it useful in distinguishing this disease. This is clearly demonstrated in Table 1. for this table was obtained as follows. Total cellular RNA was isolated from the indicated actively growing cell lines as described by Chirgwin, et al., "Isolation of biologically active ribonucleic acid from sources enriched in ribonuclease, " Biochemistry 1979; 18:5294-5299. samples were fractionated over a 1% agarose-formaldehyde gel and transferred to a nylon membrane (Qiagen, Chatsworth, CA) by capillary action. The hybridization probe was generated from a 1 kilobase pair BstXI restriction fragment isolated from pLC56, a plasmid harboring the HCAVIII gene in its initial isolation. This fragment was radiolabeled with 32P using a PRIME-IT® Random Primer Labeling Kit obtained from Stratagene, La Jolla, CA. A membrane containing RNA derived from healthy human tissue was purchased from Clonetech Laboratories, Inc., Palo Alto, CA. RNA blots were hybridized in a standard cocktail containing 32P-labeled probe at 42°C

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overnight then exposed to X-ray film. The same blots were subsequently, upon removal of the probe, rehybridized with a second ^{12}P -labeled DNA from β -actin to serve as a positive control for integrity of the blotted RNA.

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As shown in Table 1, normal lung tissue does not express the HCAVIII gene in detectable amounts. Other tumor cell lines fail to express, or express only in minor amounts, which will allow easy distinction of non-small cell carcinomas.

TABLE 1. NORTHERN BLOTS USING HCAVIII CDNA AGAINST NORMAL TISSUES AND TUMOR CELL LINES

	TISSUE	mRNA (kB)	INTENSITY
	NORMAL TISSUE		
5	heart	nd^1	
	brain	4.5	1X ²
	placenta	4.5	1X
	lung	nd	
	liver	nd	
10	skeletal muscle	nd	
	kidney	4.5	100X
	pancreas	4.5	10X
	TUMOR CELL LINE		
	A549 (lung carcinoma)	3.5	50 00X
15		5.4	5 0X
		8.0	25 X
		9.0	25X
	BT20 (breast carcinoma)	nd	
	G361 (melanoma)	nd	
20	HT144 (melanoma)	nd	
	U937 (histiocytic lymphoma)	nd	
	KG-1 (myelogenous leukemia)	nd	
25	nd = none detected		

nd = none detected

² 1X = at limit of detection

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In one embodiment of the invention, probes are made corresponding to sequences of the cDNA shown in SEQ ID NO:3, which are complimentary to the mRNA for HCAVIII. These probes can be radioactively or non-radioactively labeled in a number of ways well known to the art. The probes can be made of various lengths. Such factors as stringency and GC content may influence the desired probe length for particular applications. The probes correspond to a length of 10-986 nucleotides from SEQ ID NO:3. The labeled probes can then be bound to detect the presence or absence of mRNA encoding the HCAVIII in biopsy material through in situ hybridization. The mRNA is expected to be associated with the presence of non-small cell tumors and to be a marker for the precancerous condition as well.

In situ hybridization provides a specificity to the target tissue that is not obtainable in Northern, PCR or other probe-driven technologies. In situ hybridization permits localization of signal in mixed-tissue specimens commonly found in most tumors and is compatible with many histologic staining procedures. This technique is comprised of three basic components: first is the preparation of the tissue sample provided by the pathologist to permit successful hybridization to the probe. Second is the preparation of the hybridization probe, typically a RNA complementary to the mRNA of the gene of interest (i.e., antisense RNA). RNA probes are preferred over DNA probes for in situ hybridizations mainly because background hybridization of the probe to irrelevant nucleic acids or nonspecific attachment to cell debris or subcellular organelles can be eliminated with RNAse treatment post-hybridization. Third is the hybridization and post-hybridization detection. Typically

the RNA transcript probe has been radiolabeled by the incorporation of ³²P or ³⁵S nucleotides to permit subsequent detection of the probed specimen by autoradiography or quantitation of silver grains following treatment with autoradiographic emulsion. Nonradioactive detection systems have also been developed. In one example, biotinylated nucleotides can be substituted for the radioactive nucleotide in the RNA probe preparation, permitting visualization of the probed sample by immunocytochemistry-derived techniques. Example 1 describes in situ hybridization procedures using RNA probes derived from the HCAVIII gene. Example 2 provides exemplary fluorescent in situ (FISH) hybridization procedures.

The cDNA for HCAVIII (SEQ ID NO:3) is currently in an expression vector which is be used to generate the protein in *E. coli*. This expression system described in Example 3 produces HCAVIII to be used as an antigen for the generation of antibodies (Example 4) for use in an ELISA assay to detect shed HCAVIII in body fluids as described in Example 5. The methods for production of antibodies and ELISA type assays are well known in the art. Exemplary methods and components of these procedures have been chosen and developed and are described in Examples 4 and 5.

The expression and purification of foreign proteins in *E. coli* is often problematic. On occasion, the protein is expressed at high levels but is deposited within the cell as an insoluble, denatured form termed an inclusion body. These bodies are often observed when the foreign protein contains a hydrophobic domain, such as found in the membrane spanning segment of HCAVIII. Through

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recombinant DNA technology, the DNA sequences encoding the membrane spanning segment of HCAVIII are deleted. protein expressed in E. coli from this engineered plasmid is now in a soluble and native form within the cell, permitting a rapid and less harsh purification. addition, the ELISA test to measure HCAVIII shed into body fluids as described in Example 5 relies on the recombinant protein produced from E. coli. Typically, the shed antigen is a membrane-bound receptor that was released from the membrane spanning segment anchoring it to the cell. Consequently, the recombinant HCAVIII engineered to remove the membrane spanning segment is a more accurate representation of the putative HCAVIII shed antigen found in specimens and may prove to be the preferred antigen for polyclonal antisera and monoclonal antibody production as described for the development of an ELISA test.

To produce the engineered plasmid, a first plasmid is constructed by cleaving pLC56 with the restriction enzyme Tthl11 I, followed by treatment with T₄-DNA polymerase and dGTP, dATP, dTTP and dCTP, and finally with alkaline phosphatase to remove 5'-terminal phosphates. sample is then purified by phenol/chloroform extraction and ethanol precipitation. The sample is digested with the restriction endonuclease BspEl, then the fragments are resolved by agarose gel electrophoresis to permit the isolation of a 267 base pair fragment. A second plasmid described previously for expression of the HCAVIII mature protein (SEQ ID NO:4), is cleaved with EcoRI and BspE1 followed by alkaline phosphatase treatment and purification by phenol/chloroform extraction and ethanol precipitation. Two oligonucleotides are synthesized, being 5'-TGAGTCGACG (SEO ID NO:10) and 5'-AATTCGTCGACTCA

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(SEQ ID NO:11), that complement each other and upon annealing, provide a termination codon (TGA) and sequence complementary to EcoRI cleaved DNA. Finally, the two oligonucleotides, the 267 base pair fragment, and the BspEI/EcoRI cleaved plasmid will be combined in a ligation reaction, and the resultant plasmid which contains the truncated DNA sequence (SEQ ID NO:12) is used to transform competent E. coli. Upon expression in E. coli, the resulting truncated protein (SEQ ID NO:13) is 271 amino acids as determined by SDS polyacrylamide electrophoresis and of a size consistent with other HCA's but lacking the membrane spanning segment and the intracellular domain. A second plasmid encoding a HCAVIII truncated protein (SEQ ID NO:14) lacking the membrane spanning segment and intracellular domain was created as described above, except that restriction enzyme Ple I was substituted for Tth111 I, resulting in a gel purified DNA fragment of 276 base pairs. Upon expression in E.coli, the resulting protein is now 274 amino acids (SEQ ID NO:15).

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An understanding of protein phosphorylation and its role in the mechanism of cell transformation has been actively pursued, most notably with tyrosine phosphorylation and oncogene activation. The role of serine/threonine protein phosphorylation by a variety of protein kinases including protein kinase C has been studied extensively with respect to signal transduction, but its role in oncogenesis is less clear. To provide a valuable tool to be used in the study of the role of HCAVIII serine phosphorylation in oncogenesis, an altered cDNA can be prepared to code for an altered protein. Changes to amino acids other than "Gly" may be realized by alterations to the oligonucleotide sequence (SEQ ID NO:16)

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used to encode the selected residue. Other modifications to alter the serine phosphorylation site would utilize the described technology to modify either both "Arg" residues located within SEQ ID NO:9 or amino acid residues 299 and 300 of SEQ ID NO:2, SEQ ID NO:4 and SEQ ID NO:6. "Arg" residues contain a net positive charge, the substituted amino acids would preferably be "Lys" or "His," also positively charged amino acids. An exemplary plasmid is produced in which the "Ser" codon (amino acid residue 4 of SEQ ID NO:9; amino acid residue 302 in SEQ ID NO:2, SEQ ID NO:4 and SEQ ID NO:6), is converted to a "Gly" codon using an in vitro mutagenesis technique described in Example 3 and previously recited in Kunkel, Thomas, "Rapid and efficient site-specific mutagenesis without phenotypic selection, " Proc Natl Acad Sci USA 1985; 82:488-492, and the oligonucleotide 5'-CTTTTTTGATACCCTTCCTGAA (SEQ ID NO:16) (located in SEQ ID NO:1 at the base pairs 1010-1034 with 1022 as the mutagenized base pair). The DNA sequences containing the HCAVIII gene engineered for production of the mature protein and mutagenized codon is released from the mutagenesis vector by BamHI and EcoRI restriction endonucleases and ligated into pGEX4T1 cleaved with the same enzymes, and the resultant plasmid is used to transform competent E. coli. The codon mutagenesis is confirmed by DNA sequence analysis, and the protein is expressed and purified from E. coli as described in Example 3. The DNA sequence of the altered plasmid as shown in SEQ ID NO:17 differs from the gene encoding the mature protein (SEQ ID NO:3) in that the nucleotide 1022 is changed from "A" to "G", and the protein sequence (SEQ

ID NO:18) expressed by the altered plasmid is identical to

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the mature protein (SEQ ID NO:4) except that amino acid residue 302 is changed from "Ser" to "Gly."

Another way to detect the presence of increased HCAVIII could be to assay for levels of carbonic anhydrase activity in biopsy materials as described in Example 6. This should be a useful test as HCAVIII, although it is an immunologically unique molecule, contains small but distinct regions which are conserved between previously reported carbonic anhydrase proteins.

In another embodiment of the invention, primers are made complimentary to the HCAVIII cDNA (SEQ ID NO:3) for detecting expression of the gene. PCR amplification of cDNA from lung biopsy cells would indicate the presence of the same non-small cell lung carcinoma.

Due to the non-small cell lung cancer specificity of HCAVIII and the gene encoding the protein, antibodies specific for HCAVIII would also exhibit non-small cell lung cancer specificity which can be employed for diagnostic detection of HCAVIII in body fluids such as serum or urine or HCAVIII containing cells. Targeting of cancer therapeutic drugs to HCAVIII containing cells can also be developed using HCAVIII specific antibodies. The genetic expression of the gene encoding HCAVIII could be modulated by drugs or anti-sense technology resulting in an alteration of the cancer state of the HCAVIII containing cells.

Example 1

In Situ Hybridization using RNA Probes

Derived from the HCAVIII Gene

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Tissue samples are treated with 4% paraformaldehyde (or equivalent fixative), dehydrated in sequential ethanol solutions of increasing concentrations (e.g., 70%, 95% and 100%) with a final xylene incubation (see Current Protocols in Molecular Biology, pp. 14.01-14.3 and Immunocytochemistry II:IBRO Handbook Series: Methods in the Neurosciences Vol 14; pp 281-300, incorporated herein by reference). The tissue is embedded in molten paraffin, molded in a casting block and can be stored at room temperature. Tissue slices, typically 8 µm thick, are prepared with a microtome, dried onto gelatin-treated glass slides and stored at -20°C.

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DNA sequences from the HCAVIII gene (SEQ ID NO:3) are subcloned into a plasmid engineered for production of RNA probes. In this example, a 776 bp DNA fragment is released from a pLC56 plasmid following BamHI/AccI digestion, where the BamHI site has been created by in vitro mutagenesis (see E. coli expression below). fragment is ligated into pGEM-2 (Promega Biotec, Madison, WI) that was cleaved with BamHI and AccI and transformed into competent E. coli. This constructed plasmid contains the T7 RNA polymerase promoter downstream of the AccI restriction site and hence can drive transcription of the antisense HCAVIII sequences defined by the BamHI/AccI Following linearization of the subsequent plasmid with BamHI, an in vitro transcription reaction composed of transcription buffer (40 mM Tris-HCl, pH 7.5, 6 mM MgCl₂, 2 mM spermidine, 10 mM NaCl, 10 mM dithiothreitol, 1 U/ul ribonuclease inhibitor), linearized plasmid, 10 mM GTP, 10 mM ATP, 10 mM CTP, 100 µCi of (35S)UTP, and T7 RNA polymerase is incubated at 37°C. Multiple RNA copies of the gene are produced that then are

used as a hybridization probe. The reaction is terminated by the addition of DNAase, and the synthesized RNA is recovered from unincorporated nucleotides by phenol/chloroform extraction and sequential ethanol precipitations in the presence of 2.5 M ammonium acetate.

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The slides containing fixed, sectioned tissues are rehydrated in decreasing concentration of ethanol (100%, 70% and 50%), followed by sequential treatments with 0.2 N HCl, 2X SSC (where 20X SSC is 3 M NaCl and 0.3 M sodium citrate) at 70°C to deparaffinate the sample, phosphate buffered saline (PBS), fixation in 4% paraformaldehyde and PBS wash. The slides are blocked to prevent nonspecific binding by the sequential additions of PBS/10mM dithiothreitol (45°C), 10 mM dithiothreitol/0.19% iodoacetamide/0.12% N-ethylmaleimide and PBS wash. slides are equilibrated in 0.1M triethylamine, pH 8.0, followed by treatment in 0.1M triethylamine/0.25% acetic anhydride and 0.1 M triethylamine/0.5% acetic anhydride and washed in 2X SSC. The slides are then dehydrated in increasing concentrations of ethanol (50%, 70% and 100%) and stored at -80°C.

A hybridization mix is prepared by combining 50% deionized formamide, 0.3 M NaCl, 10 mM Tris-HCl, pH 8.0, 1 mM EDTA, 1X Denhardt's solution (0.02% Ficol1 400, 0.02% polyvinylpyrrolidone, 0.02% bovine serum albumin (BSA)), 500 μg/ml yeast tRNA, 500 μg/ml poly(A), 50 mM dithiothreitol, 10% polyethyleneglycol 6000 and the ³⁵S-labeled RNA probe. This solution is placed on the fixed, blocked tissue slides which are then incubated at 45°C in a moist chamber for 0.5 to 3 hours. The slides are washed to remove unbound probe in 50% formamide, 2X SSC, 20 mM 2-mercaptoethanol (55°C), followed by 50% formamide, 2X SSC,

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20 mM 2-mercaptoethanol and 0.5% Triton-X 100 (50°C) and finally in 2X SSC/20 mM 2-mercaptoethanol (room temperature). The slides are treated with 10 mM Tris-HCl, pH 8.0/0.3 M NaCl/40 µg/ml RNase A/2 µg/ml RNAse T1 (37°C) to reduce levels of unbound RNA probe. Following RNAse treatment, the slides are washed in formamide/SSC buffers at 50°C, room temperature and then dehydrated in increasing ethanol concentrations containing 0.3 M ammonium acetate, and one final 100% ethanol wash. The slides are then exposed to X-ray film followed by emulsion autoradiography to detect silver grains.

Test tissue samples are compared to matched controls derived from normal lung tissue. Evidence of elevated transcription of the HCAVIII gene in test tissue compared to normal tissue, as determined by autoradiography (X-ray film) or alternatively by the quantitation of silver grains following emulsion autoradiography would provide evidence of a positive diagnosis for lung cancer.

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Fluorescent In Situ Hybridization (FISH) Using DNA Probes Derived from the HCAVIII Gene

A genomic clone to the HCAVIII gene (SEQ ID NO:1) is isolated using a PCR primer pair which have been identified from the pLC56 cDNA sequence. This primer pair is located in putative exon 6 of the pLC56 gene, and they are identified as Probe Exon 6A (5'-ACATTGAAGAGCTGCTTCCGG-3'; SEQ ID NO:19) and Probe Exon 6B (5'-AATTTGCACGGGGTTTCGG-3'; SEQ ID NO:20). The genomic clone of HCAVIII is then identified as a PCR product of about 119 bp using this primer pair from the designated genomic clone. This result is confirmed by Southern blotting and

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DNA sequence analysis. A sequence of 1363 bp derived from the HCAVIII genomic clone is reported in SEQ ID NO:21. This sequence is located directly before the HCAVIII cDNA and constitutes the putative promoter of this gene and likely contains transcription regulatory elements directly implicated in HCAVIII expression.

The DNA probe comprising the genomic clone of HCAVIII plus flanking sequences is labeled in a random primer reaction with digoxigenin-11-dUTP (Boehringer Mannheim Biochemicals, Indianapolis, IN) by combining the DNA with dNTP(-TTP, final 0.05 mM), digoxigenin-11-dUTP/dTTP (0.0125 mM and 0.0375 mM, final), 10 mM 2-mercaptoethanol, 50 mM Tris-HCl, pH 7.5, 10 mM MgCl₂, 20 U of DNA polymerase I and 1 ng/ml DNAase. The reaction is incubated at 15°C for two hours, and then terminated by adding EDTA to a final concentration of 10 mM. The labeled DNA probe is further purified by gel filtration chromatography. It is apparent to those skilled in the art that other suitable substrates such as biotin-11-dUTP can be substituted for digoxigenin-11-dUTP in the procedure above.

A hybridization mix is prepared by combining 50% deionized formamide, 0.3 M NaCl, 10 mM Tris-HCl, pH 8.0, 1 mM EDTA, 1X Denhardt's solution (0.02% Ficoll 400, 0.02% polyvinylpyrrolidone, and 0.02% bovine serum albumin), 500 μ g/ml yeast tRNA, 500 μ g/ml poly(A), 50 mM dithiothreitol, 10% polyethyleneglycol 6000, and the labeled DNA probe.

Single cell suspensions of tissue biopsy material or normal tissue are fixed in methanol/glacial acetic acid (3:1 vol/vol) and dropped onto microscope slides.

(Aanastasi, et al., "Detection of Trisomy 12 in chronic lymphocytic leukemia by fluorescence in situ hybridization

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to interphase cells: a simple and sensitive method," Blood 1992; 77:2456-2462). After the slides are heated for 1-2 hours at 60°C, the hybridization mix is applied to the slides which are then incubated at 45°C in a moist chamber for 0.5-3 hours. After incubation, the slides are washed three times with a solution comprising 50% formamide and 2X SSC at 42°C, washed twice in 2X SSC at 42°C, and finally washed in 4X SSC at room temperature. The slide is blocked with a solution of 4X SSC and 1% BSA, and then washed with a solution of 4X SSC and 1% Triton X-100.

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The hybridized digoxigenin-labeled probe is detected by adding a mixture of sheep anti-digoxigenin antibody (Boehringer Mannheim) diluted in 0.1 M sodium phosphate, pH 8.0, 5% nonfat dry milk, and 0.02% sodium azide, followed by the addition of fluorescein-conjugated rabbit anti-sheep IG for detection. The slides are then washed in PBS, mounted in Vectashield (Vector Laboratories, Inc., Burlingame, CA), and viewed by fluorescent microscopy.

Hybridization signals are enumerated in tumor derived tissue and then compared to normal tissue. Normal tissue displays two distinct hybridization signal characteristics of a diploid state. Enumeration over the rate of two hybridization signals/cell is considered significant.

Example 3

Expression of HCAVIII

Expression of foreign proteins is often performed in *E. coli* when an immunogen or large amounts of protein are desired, as in the development of a diagnostic kit. A preferred system for *E. coli* expression has been described (Smith, et al., "Single-step purification of polypeptides expressed in *Escherichia coli* as fusions with glutathione-

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s-transferase," Gene 1988; 67:31-40) whereby glutathione transferase is expressed with amino acids representing the cloned protein of interest attached to the carboxylterminus. The fusion protein can then be purified via affinity chromatography and the protein of interest fused to glutathione transferase released by digestion with the protease thrombin or alternatively the fusion protein is released intact from the affinity column by competing levels of free glutathione.

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To express the HCAVIII protein (SEQ ID NO:4) of this invention in E. coli using the above described technology, an expression plasmid was produced fused to the glutathione transferase gene in frame with the HCAVIII gene (SEQ ID NO:1) to produce a fusion protein. fusion gene/expression plasmid was assembled from nucleic acids derived from the following sources. First, the expression plasmid pGEX4T1 (Pharmacia, Piscataway, NJ) was cleaved in the polycloning region with the restriction endonucleases BamHI and EcoRI to permit insertion of the HCAVIII gene. Second, an oligonucleotide was synthesized, being 5'-GTCCACTTGGATCCGTTCACTGG-3' (SEQ ID NO:22). the in vitro mutagenesis procedure described by Kunkel (Proc Natl Acad Sci USA 1985; 82:488-492) and the above oligonucleotide, a BamHI restriction site was created without altering the amino acid codons of the original protein. In addition the created BamHI site was situated in correct reading frame and proximity to the predicted cleavage site separating the signal peptide from the The DNA sequences encoding the mature mature protein. protein were released from the mutagenesis vector as a BamHI/EcoRI fragment, where the EcoRI site originates from

a polycloning region of the DNA sequencing vector pUC19

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anhydrase II was obtained.

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found downstream of the HCAVIII gene. The DNA fragments described above comprised of pGEX4T-1 cleaved at BamHI and EcoRI and the HCAVIII gene released as a BamHI/EcoRI fragment was combined in a mixture composed of 1X T4 ligase buffer (50 mM Tris-HCl, 10 mM MgCl2, 20 mM dithiothreitol, 1 mM ATP, 50 µg/ml BSA, final pH 7.5) and T, DNA ligase (New England Biolabs, Beverly, MA). ligated DNA was used to transform a suitable strain of E. coli such as XL-1 Blue (Stratagene). The recovered plasmid is sequenced to confirm the expected DNA sequence. Protein expression is induced in E. coli with the chemical isopropyl β -thiogalactoside, and the fusion protein is released by cell lysis, followed by denaturation and resolubilization of the fusion protein with 8 M urea/ 20 mM Tris.Cl (pH 8.5)/10 mM dithiothreitol, dialysis and protein renaturation, and finally binding to an affinity column composed of glutathione-agarose (Sigma, St. Louis, MO) and cleavage with thrombin to release the HCAVIII protein. The resulting protein is suitable as an immunogen for polyclonal or monoclonal antibody production and for usage in an ELISA kit as a internal standard and positive control. Carbonic anhydrase enzyme activity (as described in Example 6) was measured for E.coli-derived HCAVIII and HCAVIII-truncated form (SEQ ID NO:15) and compared to commercially obtained human carbonic anhydrase II (Sigma, St. Louis, Mo.). The activity, as reported in Enzyme Unit (U)/mg, for human carbonic anhydrase II was 3571 U/mg, for HCAVIII was 274 U/mg and HCAVIII truncated form was 2632 U/mg. These results indicated an enzymatically active and renaturable HCAVIII derived from E. coli of comparable enzymatic activity to human carbonic

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The length of the resulting protein can be varied by altering the length of SEQ ID NO:1 prior to insertion into the expression plasmid, or by cleavage of amino acids from the protein resulting in the above example. Structure/ function studies of other HCA's suggest modifications (as defined by deletions at the N-terminal and C-terminal) more extensive than disclosed in SEQ ID NO:12 would still permit the production and use of a protein as an immunogen or standard, these deletions being a protein defined by about amino acid residue 3 to amino acid residue 259 in SEQ ID NO:12. Using existing technology one could synthesize a peptide of approximately 10 to 40 amino acids in length that comprises a structural domain of HCAVIII. This synthesized peptide, coupled to a carrier protein, could be used for generating polyclonal antisera specific for native HCAVIII.

Example 4

Production of Antibodies to HCAVIII

The production of polyclonal antisera is described in great detail in Harlow, et al., Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratories, New York, 1988 incorporated herein by reference. The HCAVIII protein (SEQ ID NO:4) in the presence of an adjuvant is injected into rabbits with a series of booster shots as a prescribed schedule optimal for high titers of antibody in serum. A total of seven biweekly bleeds were obtained from two rabbits immunized with HCAVIII truncated protein (SEQ ID NO:15). The resulting anti-HCAVIII serum titer was compared to preimmune sera of the same rabbits and determined to be 1000 to 2000-fold greater, hence suitable as a reagent for indirect ELISA (Example 5). Rabbit

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antibody was partially purified by precipitation with ammonium sulfate (50%, final) followed by dialysis and fractionation by preparative DEAE-HPLC.

An extensive description for producing monoclonal antibodies derived from the spleen B cells of an immunized mouse and a immortalized myeloma cell is found in the above reference for polyclonal antisera production. Mice are immunized with either the purified HCAVIII protein or a glutathione/HCAVIII fusion protein. Following cell fusion, selection for hybrid cells and subcloning, hybridomas are screened for a positive antibody against whole A549 cells or purified HCAVIII protein using an indirect ELISA assay as described for the ELISA kit (see Example 5).

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ELISA Assay of Shed HCAVIII

An indirect ELISA screening assay for HCAVIII protein (SEQ ID NO:4) has been designed to detect and monitor the HCAVIII protein in body fluids including but not limited to serum and other biological fluids such as sputum or bronchial effluxion at effective levels necessary for sensitive but accurate determinations. It is intended to aid in the early diagnosis of non-small cell lung cancer, for which there currently is no effective treatment. An early-detection, accurate, non-invasive assay for non-small cell lung cancer would be of great benefit in the management of this disease.

The immunochemicals used in this procedure were rabbit anti-human HCAVIII antibody (purified IgG, IgM) produced according to the procedure given in Example 4, mouse anti-human HCAVIII (monoclonal) also produced

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according to the procedure given in Example 4, and goat anti-Rabbit IgG/peroxidase conjugate. The HCAVIII protein standard and internal positive control were produced as described in Example 3 for expression in *E. coli*.

Substrate components include 1 M H_2SO_4 stored at room temperature and 3',5,5'-tetramethylbenzidine (TMB) (Sigma Chemical Co.) used as a peroxidase substrate and stored at room temperature in the dark to prevent exposure to light.

Several buffers, diluents, and blocking agents were used in the procedure. Note that no sodium azide preservative was used in any of the buffers. This was done to avoid any possible interference from the azide with the peroxidase conjugate.

Phosphate buffered saline (PBS) was prepared by adding 32.0 g sodium chloride, 0.8 g potassium phosphate, monobasic, 0.8 g potassium chloride, and 4.6 g sodium phosphate, dibasic, anhydrous, to 3.2 L deionized water and mixing to dissolve. After bringing the solution to 4 L with deionized water and mixing, the pH was about 7.2. The buffer can be stored at 4°C for a maximum of 3 weeks.

Two bovine serum albumin solutions (BSA) were utilized as diluents. A 1% BSA solution in PBS, utilized as the second antibody/conjugate diluent, was prepared by adding 1 g BSA (bovine albumin, Fraction V, Sigma Chemical Co.) to 80 ml of PBS, allowing it to stand as it slowly goes into solution, adding PBS to a final volume of 100 ml, and then mixing. This diluent can be stored at 4°C for a maximum of 2 weeks; however if the solution becomes turbid, it should be discarded. As a diluent for the standards and samples, a 0.025% BSA solution in PBS was prepared fresh for each assay by diluting the 1% BSA diluent with PBS 1:40 (vol/vol).

A borate blocking buffer (0.17 M H₃BO₃, 0.12 M NaCl, 0.05% Tween 20, lmM EDTA and 0.25% BSA was also used.

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The substrate buffer was phosphate-citrate/sodium per borate (Sigma, St. Louis, Mo.).

All assays were performed in Immulon IV plates (Dynatech, Chantilly, VA #011-010-6301). The assay plates were coated with a monoclonal antibody against HCAVIII by adding 50 ul of a 10 ug/ml solution of antibody in PBS to each well of Immulon IV plates. The plates were covered and incubated overnight at room temperature. The antibody solution was removed and the wells rinsed three times with deionized water. Three-hundred microliters (300 ul) of the borate blocking buffer was added to each well and incubated at room temperature for thirty minutes. The buffer was removed, the wells rinsed three times with deionized water, and the plates air dried. The plates were then wrapped and stored at 4°C.

The standard E.coli-derived HCAVIII truncated protein (SEQ ID NO:15), was diluted to 32 ng/ml in PBS/0.025% BSA and two-fold serial dilutions were made in same. The samples were also diluted in PBS/0.025% BSA and 50 ul of standard or sample was applied to each well. The plates were incubated overnight, covered, at room temperature.

The standard and sample solutions were removed from the wells and the wells were rinsed three times with deionized water. Three-hundred microliters (300 ul) borate blocking buffer was added to each well and incubated at room temperature for thirty minutes. The plates were rinsed again with deionized water and tapped (inverted) on paper towels to remove excess water. The second antibody rabbit antisera to HCAVIII truncated protein (SEQ ID NO:15), was diluted to 1 ug/ml in PBS/1%

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BSA and 50 ul was added to each well. The plates were covered and incubated at room temperature two hours.

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The antibody solution was removed from the wells which were then rinsed with deionized water three times. They were then blocked for ten minutes at room temperature with borate blocking buffer, rinsed again with deionied water three times, and tapped on paper towels. The antibody conjugate, goat F(ab')2 x rabbit IgG & IgL-HPRO (Tago, Camarillo, CA.) was diluted 1:16,000 in PBS/1%BSA and 50 ul was added to each well. The plates were covered and incubated at room temperature two hours.

The antibody conjugate solution was removed from the wells and they were rinsed with deionized water three times, blocked with three-hundred ul borate buffer at room temperature then minutes, rinsed three times with deionized water, and tapped on paper towels. substrate was prepared no more than fifteen minues before use by dissolving one capsule of phosphate-citrate/sodium perborate (Signma, St. Louis, Mo.) in 100 ml water. each plate, one tablet of TMB was added to 10 ml of the phosphate-citrate/sodium perborate buffer and syringe filtered. One-hundred ul was added to each well and the plates were covered and incubated at room temperature in the dark for one hour. The reaction was stopped by adding 50 ul of 1M H₂SO₄ to each well. The plates were read on a Molecular Devices microplate reader at 450nm. Under these conditions, a linear response was obtained from 0.5 to 32 ng/ml using HCAVIII truncated protein as a standard, with the assay sensitivity at 0.5 ng/ml. No cross-reaction was observed against HCAII, an abundant carbonic anhydrase in human serum.

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Example 6

Carbonic Anhydrase (CA) Activity of Biopsy Tissue

Ice cold solutions of ITB (20 mM imidazole, 5 mM Tris, and 0.4 mM para-nitrophenol, pH 9.4-9.9) and Buffer A (25 mM triethanolamine, 59 mM $\rm H_2SO_4$, and 1 mM benzamidine HCl) are prepared.

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A homogenate is prepared by scraping with a cell scraper into 1-2 ml of Buffer A a monolayer of tissue cells cultured from a tissue sample taken from a biopsy. A portion of the sample is then boiled to inactivate CA.

A tube is placed in an ice water bath. For the macroassay, a 10 x 75 mm glass tubes and rubber stopper with 16 gauge and 18 gauge needle ports is used; for the microassay, a 6 x 50 mm glass tubes and rubber stopper with 18 gauge needle port and 20 gauge needle with attached PE90 tubing. The sample is added and along with ice cold water to a final volume of 500 µl for macroassay or 50 µl for microassay. 500 µl (macro) or 50 µl (micro) ice cold water is used for a water control. 10 µl antifoam (A. H. Thomas, Philadelphia, PA) is added to the tube which is then incubated in ice water for 0.5 to 3 minutes.

The tube is capped with a stopper and CO_2 at 150 ml/min (macro) or 100 ml/min (micro) is bubbled through the smaller needle port for 30 sec.

50 µl (macro) or 50 µl (micro) of the ITB solution is rapidly added through the larger needle port with a cold Hamilton syringe. The sample becomes yellow.

Using a timer or stopwatch, the time at which the solution in the tube becomes colorless is measured and recorded. The tube may be momentarily removed from the bath and held in front of a white background to determine

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the color change. Comparison to a previously acidified sample may be used.

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The procedure is repeated with the boiled sample.

The volume of sample that corresponds to approximately one enzyme unit is determined using the formula below.

Volume (1EU) = V_{EU} = volume used x log2/log (boiled time/activated time) One enzyme unit is the activity that halves the boiled control time.

The assay is repeated 1-3 times with the sample and boiled sample, using the adjusted volume of sample.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT:
 - (A) NAME: Cytoclonal Pharmaceutics, Inc.
 - (B) STREET: 9000 Harry Hines Blvd, Suite 330
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 - (H) TELEFAX: (214) 350-9514
 - (I) TELEX:
- (ii) TITLE OF INVENTION: Lung Cancer Marker
- (iii) NUMBER OF SEQUENCES: 22
- (iv) CORRESPONDENCE ADDRESS:
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 - (D) STATE: TX
 - (E) COUNTRY: US
 - (F) ZIP: 75270-2197
 - (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.30
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
 - (A) NAME: John A. Harre
 - (B) REGISTRATION NUMBER: 37,345
 - (C) REFERENCE/DOCKET NUMBER: B35792CIPPCT
 - (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 214-939-4500
 - (B) TELEFAX: 214-939-4600
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1104 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: both
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

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(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 321093	
(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 1191093	
<pre>(ix) FEATURE:</pre>	
GCCCGCGCCC GCCCCGCAGG AGCCCGCGAA G ATG CCC CGG CGC AGC CTG CAC Met Pro Arg Arg Ser Leu His -29 -25	52
GCG GCG GCC GTG CTC CTG GTG ATC TTA AAG GAA CAG CCT TCC AGC Ala Ala Ala Val Leu Leu Val Ile Leu Lys Glu Gln Pro Ser Ser -20 -15 -10	100
CCG GCC CCA GTG AAC GGT TCC AAG TGG ACT TAT TTT GGT CCT GAT GGG Pro Ala Pro Val Asn Gly Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly -5 1 5 10	148
GAG AAT AGC TGG TCC AAG AAG TAC CCG TCG TGT GGG GGC CTG CTG CAG Glu Asn Ser Trp Ser Lys Lys Tyr Pro Ser Cys Gly Gly Leu Leu Gln 15 20 25	196
TCC CCC ATA GAC CTG CAC AGT GAC ATC CTC CAG TAT GAC GCC AGC CTC Ser Pro Ile Asp Leu His Ser Asp Ile Leu Gln Tyr Asp Ala Ser Leu 30 35 40	244
ACG CCC CTC GAG TTC CAA GGC TAC AAT CTG TCT GCC AAC AAG CAG TTT Thr Pro Leu Glu Phe Gln Gly Tyr Asn Leu Ser Ala Asn Lys Gln Phe 45 50 55	292
CTC CTG ACC AAC AAT GGC CAT TCA GTG AAG CTG AAC CTG CCC TCG GAC Leu Leu Thr Asn Asn Gly His Ser Val Lys Leu Asn Leu Pro Ser Asp 60 65 70	340
ATG CAC ATC CAG GGC CTC CAG TCT CGC TAC AGT GCC ACG CAG CTG CAC Met His Ile Gln Gly Leu Gln Ser Arg Tyr Ser Ala Thr Gln Leu His 75 80 85 90	388

CTG CAC TGG GGG AAC CCG AAT GAC CCG CAC GGC TCT GAG CAC ACC GTC

Leu His Trp Gly Asn Pro Asn Asp Pro His Gly Ser Glu His Thr Val 95 100 105

AGC GGA CAG CAC TTC GCC GCC GAG CTG CAC ATT GTC CAT TAT AAC TCA

Ser Gly Gln His Phe Ala Ala Glu Leu His Ile Val His Tyr Asn Ser 110 115 120

110

GAC As p	CTT Leu	TAT Tyr 125	CCT Pro	GAC Asp	GCC Ala	AGC Ser	ACT Thr 130	GCC Ala	AGC Ser	AAC Asn	AAG Lys	TCA Ser 135	GAA Glu	GGC Gly	CTC Leu	532
														TCC Ser		580
GAC Asp 155	AAG Lys	ATC Ile	TTC Phe	AGT Ser	CAC His 160	CTT Leu	CAA Gln	CAT His	GTA Val	AAG Lys 165	TAC Tyr	AAA Lys	GGC Gly	CAG Gln	GAA Glu 170	628
														AGG Arg 185		676
														TGC Cys		724
														TCC Ser		772
														ATG Met		820
														CAG Gln		868
														GTC Val 265		916
														CTG Leu		964
														CTT Leu		1012
		Lys												TAC Tyr		1060
	Ala				GAG Glu 32 0	Thr					TGA	GGTC	ccc	G		1104

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 354 amino acids
(B) TYPE: amino acid

(D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Pro Arg Arg Ser Leu His Ala Ala Ala Val Leu Leu Val Ile -29 -25 -20 -15

Leu Lys Glu Gln Pro Ser Ser Pro Ala Pro Val Asn Gly Ser Lys Trp
-10 -5

Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys Lys Tyr Pro 5 10 15

Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu His Ser Asp Ile 20 25 30 35

Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln Gly Tyr Asn
40 45 50

Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly His Ser Val 55 60 65

Lys Leu Asn Leu Pro Ser Asp Met His Ile Gln Gly Leu Gln Ser Arg
70 75 80

Tyr Ser Ala Thr Gln Leu His Leu His Trp Gly Asn Pro Asn Asp Pro 85 90 95

His Gly Ser Glu His Thr Val Ser Gly Gln His Phe Ala Ala Glu Leu 100 105 110 115

His Ile Val His Tyr Asn Ser Asp Leu Tyr Pro Asp Ala Ser Thr Ala 120 125 130

Ser Asn Lys Ser Glu Gly Leu Ala Val Leu Ala Val Leu Ile Glu Met 135 140 145

Gly Ser Phe Asn Pro Ser Tyr Asp Lys Ile Phe Ser His Leu Gln His 150 155 160

Val Lys Tyr Lys Gly Gln Glu Ala Phe Val Pro Gly Phe Asn Ile Glu 165 170 175

Glu Leu Leu Pro Glu Arg Thr Ala Glu Tyr Tyr Arg Tyr Arg Gly Ser 180 185 190 195

Leu Thr Thr Pro Pro Cys Asn Pro Thr Val Leu Trp Thr Val Phe Arg 200 205 210

Asn Pro Val Gln Ile Ser Gln Glu Gln Leu Leu Ala Leu Glu Thr Ala 215 220 225

Leu Tyr Cys Thr His Met Asp Asp Pro Ser Pro Arg Glu Met Ile Asn 230 235 240

Asn	Phe 245	Arg	Gln	Val	Gln	Lys 250	Phe	As p	Glu	Arg	Le u 25 5	Val	Tyr	Thr	Ser		
Phe 260	Ser	Gln	Val	Gln	Val 265	Cys	Thr	Ala	Ala	Gly 270	Leu	Ser	Leu	Gly	11e 275		
Ile	Leu	Ser	Leu	Ala 280	Leu	Ala	Gly	Ile	Leu 285	Gly	Ile	Суз	Ile	Val 290	Val		
Val	Val	Ser	11e 295	Trp	Leu	Phe	Arg	Arg 300	Lys	Ser	Ile	Lys	Lys 305	Gly	Asp		
Asn	Lys	Gly 310	Val	Ile	Tyr	Lys	Pro 315	Ala	Thr	Lys	Met	Glu 320	Thr	Glu	Ala		
His	Ala 325																
(2)	INFO	RMAT	NOIT	FOR	SEQ	ID 1	10:3:	:									
	(i)	() ()	A) LI 3) Ti 3) Si	CE CHENGTHE PER : CRANICO POLO	i: 96 nucl	86 ba Leic ESS:	se p acid both	eir:	3								
	(ii)	MOI	ECUI	LE TY	PE:	CDN											
	(ix)	(3		e: Ame/i Ocati			75										
re	•	() () ()	3) L(E: AME/I CATI THER prot	ION:	895.	. 906. : NOI!	5 : /no	ote=				ation	n sit	:e		
	(x i)	SEC	DUEN	CE DI	ESCRI	PTIC	ON: S	BEQ I	D N	0:3:							
				TAT Tyr 5												48	
				TGT Cys												96	
				CAG Gln												144	
				TCT Ser												192	

					AAC Asn 70											240
					GCC Ala			_								288
AAT Asn	GAC Asp	CCG Pro	CAC His 100	GGC G1 y	TCT Ser	GAG Glu	CAC His	ACC Thr 105	GTC Val	AGC Ser	G) y	CAG Gln	CAC His 110	TTC Phe	GCC Ala	336
GCC Ala	GAG Glu	CTG Leu 115	CAC His	ATT Ile	GTC Val	CAT His	TAT Tyr 120	AAC Asn	TCA Ser	GAC Asp	CTT Leu	TAT Tyr 125	CCT Pro	GAC Asp	GCC Ala	384
					AAG Lys											432
ATT Ile 145	GAG Glu	ATG Met	GGC Gly	TCC Ser	TTC Phe 150	AAT Asn	CCG Pro	TCC Ser	TAT Tyr	GAC Asp 155	AAG Lys	ATC Ile	TTC Phe	AGT Ser	CAC His 160	480
					TAC Tyr											528
AAC Asn	ATT Ile	GAA Glu	GAG Glu 180	CTG Leu	CTT Leu	CCG Pro	GAG Glu	AGG Arg 185	ACC Thr	GCT Ala	GAA Glu	TAT Tyr	TAC Tyr 190	CGC Arg	TAC Tyr	576
					ACA Thr											624
					GTG Val											672
					TGC Cys 230											720
					CGG Arg										GTA Val	768
															AGT Ser	816
			Ile		TCA Ser										TGT Cys	864

ATT GTG GTG GTG GTG TCC ATT TGG CTT TTC AGA AGG AAG AGT ATC AAA

11e Val Val Val Val Ser 11e Trp Leu Phe Arg Arg Lys Ser 11e Lys
290

AAA GGT GAT AAC AAG GGA GTC ATT TAC AAG CCA GCC ACC AAG ATG GAG
Lys Gly Asp Asn Lys Gly Val 11e Tyr Lys Pro Ala Thr Lys Met Glu
310

ACT GAG GCC CAC GCT TGAGGTCCCC G

986

(2) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 325 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys
1 5 10 15

Lys Tyr Pro Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu His 20 25 30

Ser Asp Ile Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln 35 40

Gly Tyr Asn Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly 50 55 60

His Ser Val Lys Leu Asn Leu Pro Ser Asp Met His Ile Gln Gly Leu 65 70 75 80

Gln Ser Arg Tyr Ser Ala Thr Gln Leu His Leu His Trp Gly Asn Pro 85 90 95

Asn Asp Pro His Gly Ser Glu His Thr Val Ser Gly Gln His Phe Ala 100 105 110

Ala Glu Leu His Ile Val His Tyr Asn Ser Asp Leu Tyr Pro Asp Ala 115 120 125

Ser Thr Ala Ser Asn Lys Ser Glu Gly Leu Ala Val Leu Ala Val Leu 130 135

Ile Glu Met Gly Ser Phe Asn Pro Ser Tyr Asp Lys Ile Phe Ser His 145 150 155 160

Leu Gln His Val Lys Tyr Lys Gly Gln Glu Ala Phe Val Pro Gly Phe 165 170 175

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60

118

Met -29

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									47						
Asn	Ile	Glu	Glu 180	Leu	Leu	Pro	Glu	Arg 185	Thr	Ala	Glu	Tyr	Tyr 190	Arg	Tyr
Arg	Gly	Ser 195	Leu	Thr	Thr	Pro	Pro 200	Cys	Asn	Pro	Thr	Val 205	Leu	Trp	Thr
Val	Phe 210	Arg	Asn	Pro	Val	Gln 215	Ile	Ser	Gln	Glu	Gln 220	Leu	Leu	Ala	Leu
Glu 225	Thr	Ala	Leu	Tyr	Cys 230	Thr	His	Met	Asp	As p 235	Pro	Ser	Pro	Arg	Glu 240
Met	Ile	Asn	Asn	Phe 245	Arg	Gln	Val	Gln	Lys 250	Phe	Asp	Glu	Arg	Leu 255	Val
Tyr	Thr	Ser	Phe 260	Ser	Gln	Val	Gln	Val 265	Cys	Thr	Ala	Ala	Gly 270	Leu	Ser
Leu	Gly	11e 275	Ile	Leu	Ser	Leu	Ala 280	Leu	Ala	Gly	Ile	Leu 285	Gly	Ile	Суѕ
Ile	Val 290	Val	Val	Val	Ser	11e 295	Trp	Leu	Phe	Arg	A rg 300	Lys	Ser	Ile	Lys
Lys 305	Gly	Asp	As n	Lys	Gly 310	Val	Ile	Tyr	Lys	Pro 315	Ala	Thr	Lys	Met	Glu 32 0
Thr	Glu	Ala	His	Ala 325											
(2)	INF	ORMA!	NOI	FOR	SEQ	ID P	NO:5	•							
	(1)	() (1	A) LI B) T) C) S1	engti (Pe : [ran]	HARAC H: 2: nuc: DEDNI DGY:	134 h Leic ESS:	acio boti	pai:	rs						
	(ii	MO:	LECUI	LE T	YPE:	CDNU	A.								
	(ix		A) N	AME/	KEY: ION:		11'	77							
	(ix		A) N	AME/	KEY: ION:										
	(xi) SE	QUEN	CE D	ESCR:	IPTI	ON:	SEQ :	ID N	0:5:					

GTACTCGCCA CGGCACCCAG GCTGCGCGCA CGCGGTCCCG GTGTGCAGCT GGAGAGCGAG

CCC CGG CGC AGC CTG CAC GCG GCC GTG CTC CTG CTG GTG ATC TTA 166

Pro	Arg	Arg	Ser -25	Leu	His	Ala	Ala	Ala -20	Val	Leu	Leu	Leu	Val -15	Ile	Leu	
					AGC Ser											214
					GGG Gly 10											262
					CAG Gln											310
					CTC Leu											358
					TTT Phe											406
					GAC Asp											454
					CAC His 90											502
					GTC Val											550
					TCA Ser											598
					CTC Leu											646
					TAT Tyr											694
					GAA Glu 170											742
					ACC Thr											790
					AAC Asn											838

Pro	GTG Val	Gln 215	ATT Ile	TCC Ser	CAG Gln	GAG Glu	Gln 220	CTG Leu	CTG Leu	GCT Ala	TTG Leu	GAG Glu 225	Thr	GCC Ala	Leu	880
TAC Tyr	TGC Cys 230	ACA Thr	CAC His	ATG Met	GAC Asp	GAC Asp 235	CCT Pro	TCC Ser	CCC Pro	AGA Arg	GAA Glu 240	ATG Met	ATC Ile	AAC Asn	AAC Asn	934
TTC Phe 245	CGG Arg	CAG Gln	GTC Val	CAG Gln	AAG Lys 250	TTC Phe	gat Asp	GAG Glu	AGG Arg	CTG Leu 255	GTA Val	TAC Tyr	ACC Thr	TCC Ser	TTC Phe 260	982
TCC Ser	CAA Gln	GTG Val	CAA Gln	GTC Val 265	TGT Cys	ACT Thr	GCG Ala	GCA Ala	GGA G1 y 270	CTG Leu	AGT Ser	CTG Leu	GGC Gly	ATC Ile 275	ATC Ile	1030
CTC Leu	TCA Ser	CTG Leu	GCC Ala 280	CTG Leu	GCT Ala	GGC Gly	ATT Ile	CTT Leu 285	GGC G1 y	ATC Ile	TGT Cys	ATT Ile	GTG Val 290	GTG Val	GTG Val	1078
GTG Val	TCC Ser	ATT 11e 295	TGG Trp	CTT Leu	TTC Phe	AGA Arg	AGG Arg 300	AAG Lys	AGT Ser	ATC Ile	AAA Lys	AAA Lys 305	GGT Gly	gat Asp	AAC Asn	1126
AAG Lys	GGA Gly 310	GTC Val	ATT Ile	TAC Tyr	AAG Lys	CCA Pro 315	GCC Ala	ACC Thr	AAG Lys	ATG Met	GAG Glu 320	ACT Thr	GAG Glu	GCC Ala	CAC His	1174
GCT Ala 325	TGA	GTC	ccc (GGAG	CTCC	ce ex	SCAC	atcci	A GG/	VAGG!	ACCT	TGC	TTG	GAC		1227
CCT	CAC	ACT 1	TCGG	CTCT	CT G	GACA	TTG	GA(CACC	CAA	GGT	GTTC:	TCT (GTAG	CTCAAT	1287
CTG	CAAA	CAT	GCCA	GGCC.	rc a	GGGA?	CCT	TG	CTGG	GTGC	CTC	CTTG	CCT :	recei	ACCATG	1347
ecc)	ACCC	CAG A	AGCCI	ATCC	GA T	CGAT	GAT(G GG	ATGC	ACTC	TCA	SACC	AAG (CAGC	AGGAAT	1407
TCA	AAGC:	rgc '	TTGC	TGTA	AC T	GTGT	SAGA:	T TG	IGAA	GTGG	TCT	GAAT'	TCT	GGAA'	TCACAA	1467
ACC	NAGC!	CAT	GCTG	GTGG	GC C	ATTA	ATGG'	T TG	GAAA	ACAC	TTT	CATC	CGG	GGCT	TTGCCA	1527
GAG	CGTG	CTT	TCAA	GTGT	CC T	GGAA	ATTC'	T GC	TGCT'	TCTC	CAA	GCTT	TCA	GACA	AGAATG	1587
TGC	ACTC	TCT ·	GCTT	AGGT'	TT T	GC T T	GGGA	A AC	TCAA	CTTC	TTT	CCTC	TGG .	AGAC	GGGGCA	1647
TCT	CCCT	CTG .	TTTA	CCTT	CT G	CTAT	GACA	A AA	CCTT	TAAT	CTG	CACC	ATT	CAAC	TCGGGG	1707
ACA	AATG	GGG .	ACAG	GAAG	GA T	CAAG	TTGT.	A GA	GAGA	AAAA	GAA	AACA	AGA	GATA	TACATT	1767
GTG	ATAT.	ATT	AGGG	ACAC	тт т	CACA	GTCC	T GT	CCTC	TGGA	TCA	CAGA	CAC	TGCA	CAGACC	1827
TTA	GGGA	ATG	GCAG	GTTC	AA G	TTCC	actt	C TT	GGTG	GGGA	TGA	GAAG	GGA	GAGA	GAGCTA	1887
GAG	GGAC	AAA	GAGA	ATGA	GA A	GACA	TGGA	T GA	TCTG	GGAG	AGT	CTCA	CTT	TGGA	ATCAGA	1947
ATT	GGAA	TCA	CATT	CTGT	TT A	TCAA	GCCA	AA T	TGTA	AGGA	CAG	AATA	ATA	CART	ATTAAG	2007

TCCAAATCCA	ACCTCCTGTC	AGTGGAGCAG	TTATGTTTTA	TACTCTACAG	ATTTTACAAA	2067
TAATGAGGCT	GTTCCTTGAA	AATGTGTTGT	TGCTGTGTCC	TGGAGGAGAC	ATGAGTTCCG	2127
AGATGAC						2134

(2) INFORMATION FOR SEQ ID NO:6:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 354 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Pro Arg Arg Ser Leu His Ala Ala Ala Val Leu Leu Val Ile -29 -25 -20 -15

Leu Lys Glu Gln Pro Ser Ser Pro Ala Pro Val Asn Gly Ser Lys Trp
-10 -5

Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys Lys Tyr Pro
5 10 15

Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu His Ser Asp Ile 20 25 30 35

Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln Gly Tyr Asn
40 45 50

Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly His Ser Val
55 60 65

Lys Leu Asn Leu Pro Ser Asp Met His Ile Gln Gly Leu Gln Ser Arg
70 75 80

Tyr Ser Ala Thr Gln Leu His Leu His Trp Gly Asn Pro Asn Asp Pro 85 90 95

His Gly Ser Glu His Thr Val Ser Gly Gln His Phe Ala Ala Glu Leu 100 105 110 115

His Ile Val His Tyr Asn Ser Asp Leu Tyr Pro Asp Ala Ser Thr Ala 120 125 130

Ser Asn Lys Ser Glu Gly Leu Ala Val Leu Ala Val Leu Ile Glu Met 135 140 145

Gly Ser Phe Asn Pro Ser Tyr Asp Lys Ile Phe Ser His Leu Gln His 150 155 160

Val Lys Tyr Lys Gly Gln Glu Ala Phe Val Pro Gly Phe Asn Ile Glu 165 170 175 WO 96/02552

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Glu 180	Leu	Leu	Pro	Glu	Arg 185	Thr	Ala	Glu	Tyr	Tyr 190	Arg	Tyr	Arg	G1 y	Se r 195
Leu	Thr	Thr	Pro	Pro 200	Cys	As n	Pro	Thr	Val 205	Leu	Trp	Thr	Val	Phe 210	Arg
As n	Pro	Val	Gln 215	Ile	Ser	Gln	Glu	Gln 220	Leu	Leu	Ala	Leu	Glu 225	Thr	Ala
Leu	Tyr	Cys 230	Thr	His	Met	Asp	Asp 235	Pro	Ser	Pro	Arg	Glu 240	Met	Ile	As n
Asn	Phe 245	Arg	Gln	Val	Gln	Lys 250	Phe	Asp	Glu	Arg	Leu 255	Val	Tyr	Thr	Ser
Phe 260	Ser	Gln	Val	Gln	Val 265	Сув	Thr	Ala	Ala	Gly 270	Leu	Ser	Leu	Gly	11e 275
Ile	Leu	Ser		Ala 280	Leu	Ala	Gly	Ile	Leu 285	Gly	Ile	Cys	Ile	Val 290	Val
Val	Val	Ser	Ile 295	Trp	Leu	Phe	Arg	Arg 300	Lys	Ser	Ile	Lys	Lys 305	Gly	As p
Asn	Lys	Gly 310	Val	Ile	Tyr	Lys	Pro 315	Ala	Thr	Lys	Met	Glu 320	Thr	Glu	Ala
His	Ala 325														

(2) INFORMATION FOR SEQ ID NO:7:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 624 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: both
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

	CCAATCTGCC	TTTGAATCTG	GAGGAAATAG	GCAGAAACAA	AATGACTGTA	GAACTTATTC	60
•	TCTGTAGGCC	AAATTTCATT	TCAGCCACTT	CTGCAGGATC	CCTACTGCCA	ACCTGGAATG	120
	GAGACTTTTA	TCTACTTCTC	TCTCTCTGAA	GATGTCAAAT	CGTGGTTTAG	ATCAAATATA	180
	TTTCAAGCTA	TAAAAGCAGG	AGGTTATCTG	TGCAGGGGGC	TGGCATCATG	TATTTAGGGG	240
	CAAGTAATAA	TGGAATGCTA	CTAAGATACT	CCATATTCTT	CCCCGAATCA	CACAGACAGT	300
	TTCTGACAGG	CGCAACTCCT	CCATTTTCCT	CCCGCAGGTG	AGAACCCTGT	GGAGATGAGT	360
	CAGTGCCATG	ACTGAGAAGG	AACCGACCCC	TAGTTGAGAG	CACCTTGCAG	TTCCCCGAGA	420
	ACTITCTGAT	TCACAGTCTC	ATTTTGACAG	CATGAAATGT	CCTCTTGAAG	CATAGCTTTT	480

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52	
TARATATOTT TTTCCTTCTA CTCCTCCCTC TGACTCTAAG AATTCTCTCT TCTGGAATCG	540
CTTGAACCCA GGAGGCGGAG GTTGCAGTAA GCCAAGGTCA TGCCACTGCA CTCTAGCCTG	600
GGTGACAGAG CGAGACTCCA TCTC	624
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 12 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: both (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: cDNA	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 112	
(x1) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
AGA AGG AAG AGT Arg Arg Lys Ser	12
(2) INFORMATION FOR SEQ ID NO:9:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 4 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: protein	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:	
Arg Arg Lys Ser	
(2) INFORMATION FOR SEQ ID NO:10:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 10 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:	
TGAGTCGACG	10
(2) INFORMATION FOR SEQ ID NO:11:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 14 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(iii) HYPOTHETICAL: NO	
(iv) ANTI-SENSE: NO	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:	
AATTCGTCGA CTCA	14
(2) INFORMATION FOR SEQ ID NO:12: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 813 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: both (D) TOPOLOGY: linear (ii) MOLECULE TYPE: CDNA	
(ix) FEATURE:	
(A) NAME/KEY: CDS (B) LOCATION: 1813	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
TCC AAG TGG ACT TAT TTT GGT CCT GAT GGG GAG AAT AGC TGG TCC AAG Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys 1 5 10 15	48
AAG TAC CCG TCG TGT GGG GGC CTG CTG CAG TCC CCC ATA GAC CTG CAC	96
Lys Tyr Pro Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu Ris 20 25 30	
AGT GAC ATC CTC CAG TAT GAC GCC AGC CTC ACG CCC CTC GAG TTC CAA Ser Asp Ile Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln 35 40 45	144
GGC TAC AAT CTG TCT GCC AAC AAG CAG TTT CTC CTG ACC AAC AAT GGC	192
Gly Tyr Asn Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly 50 55 60	

CAT His 65	TCA Ser	GTG Val	AAG Lys	CTG Leu	AAC Asn 70	CTG Leu	CCC Pro	TCG Ser	GAC Asp	ATG Met 75	CAC His	ATC Ile	CAG Gln	GGC Gly	CTC Leu 80	240
CAG Gln	TCT Ser	CGC Arg	TAC Tyr	AGT Ser 85	GCC Ala	ACG Thr	CAG Gln	CTG Leu	CAC His 90	CTG Le u	CAC His	TGG Trp	GGG Gly	AAC Asn 95	CCG Pro	208
										AGC Ser						336
GCC Ala	GAG Glu	CTG Leu 115	CAC His	ATT Ile	GTC Val	CAT His	TAT Tyr 120	AAC Asn	TCA Ser	GAC Asp	CTT Leu	TAT Tyr 125	CCT Pro	GAC Asp	GCC Ala	384
AGC Ser	ACT Thr 130	GCC Ala	AGC Ser	AAC Asn	AAG Lys	TCA Ser 135	GAA Glu	GGC Gly	CTC Leu	GCT Ala	GTC Val 140	CTG Leu	GCT Ala	GTT Val	CTC Leu	432
ATT Ile 145	GAG Glu	ATG Met	GGC Gly	TCC Ser	TTC Phe 150	AAT Asn	CCG Pro	TCC Ser	TAT Tyr	GAC Asp 155	AAG Lys	ATC Ile	TTC Phe	AGT Ser	CAC His 160	480
CTT Leu	CAA Gln	CAT His	GTA Val	AAG Lys 165	TAC Tyr	AAA Lys	GGC Gly	CAG Gln	GAA Glu 170	GCA Ala	TTC Phe	GTC Val	CCG Pro	GGA Gly 175	TTC Phe	528
AAC Asn	ATT Ile	GAA Glu	GAG Glu 180	CTG Leu	CTT Leu	CCG Pro	GAG Glu	AGG Arg 185	ACC Thr	GCT Ala	GAA Glu	TAT Tyr	TAC Tyr 190	CGC Arg	TAC Tyr	576
CGG Arg	GGG Gly	TCC Ser 195	CTG Leu	ACC Thr	ACA Thr	CCC Pro	CCT Pro 200	TGC Cys	AAC Asn	CCC Pro	ACT Thr	GTG Val 205	CTC Leu	TGG Trp	ACA Thr	624
GTT Val	TTC Phe 210	CGA Arg	AAC Asn	CCC Pro	GTG Val	CAA Gln 215	ATT Ile	TCC Ser	CAG Gln	GAG Glu	CAG Gln 220	CTG Leu	CTG Leu	GCT Ala	TTG Leu	672
GAG Glu 225	ACA Thr	GCC Ala	CTG Leu	TAC Tyr	TGC Cys 230	ACA Thr	CAC His	ATG Met	GAC Asp	GAC Asp 235	CCT Pro	TCC Ser	CCC Pro	AGA Arg	GAA Glu 240	720
ATG Met	ATC Ile	AAC Asn	AAC Asn	TTC Phe 245	CGG Arg	CAG Gln	GTC Val	CAG Gln	AAG Lys 250	TTC Phe	GAT As p	GAG Glu	AGG Arg	CTG Leu 255	GTA Val	768
TAC Tyr	ACC Thr	TCC Ser	TTC Phe 260	TCC Ser	CAA Gln	GTG Val	CAA Gln	GTC Val 265	TGT Cys	ACT Thr	GCG Ala	GCA Ala	GGA Gly 270	CTG Leu		 813

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 271 amino acids
(B) TYPE: amino acid

- (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys
1 5 10 15

Lys Tyr Pro Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu His 20 25 30

Ser Asp Ile Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln 35 40 45

Gly Tyr Asn Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly 50 55 60

His Ser Val Lys Leu Asn Leu Pro Ser Asp Met His Ile Gln Gly Leu 65 70 75 80

Gln Ser Arg Tyr Ser Ala Thr Gln Leu His Leu His Trp Gly Asn Pro 85 90 95

Asn Asp Pro His Gly Ser Glu His Thr Val Ser Gly Gln His Phe Ala 100 105 110

Ala Glu Leu His Ile Val His Tyr Asn Ser Asp Leu Tyr Pro Asp Ala 115 120 125

Ser Thr Ala Ser Asn Lys Ser Glu Gly Leu Ala Val Leu Ala Val Leu 130 135 140

Ile Glu Met Gly Ser Phe Asn Pro Ser Tyr Asp Lys Ile Phe Ser His 145 150 155 160

Leu Gln His Val Lys Tyr Lys Gly Gln Glu Ala Phe Val Pro Gly Phe 165 170 175

Asn Ile Glu Glu Leu Leu Pro Glu Arg Thr Ala Glu Tyr Tyr Arg Tyr 180 185 190

Arg Gly Ser Leu Thr Thr Pro Pro Cys Asn Pro Thr Val Leu Trp Thr 195 200 205

Val Phe Arg Asn Pro Val Gln Ile Ser Gln Glu Gln Leu Leu Ala Leu 210 215 220

Glu Thr Ala Leu Tyr Cys Thr His Met Asp Asp Pro Ser Pro Arg Glu 225 230 235 240

Met Ile Asn Asn Phe Arg Gln Val Gln Lys Phe Asp Glu Arg Leu Val 245 250 255

Tyr Thr Ser Phe Ser Gln Val Gln Val Cys Thr Ala Ala Gly Leu 260 265 270

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 822 base pairs

(B) TYPE: nucleic acid (C) STRANDEDNESS: both (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..822

(xi)	SEQUENCE	DESCRIPTION:	SEQ	ID	NO:1	4 :
------	----------	--------------	-----	----	------	-----

			GGT Gly					48
			GGC Gly					96
			GAC Asp					144
			AAC Asn 55					192
			CTG Leu					240
			ACG Thr					288
			GAG Glu					336
			CAT His					384
			TCA Ser 135					432
			AAT Asn					480

			AAA Lys					528
			CCG Pro					576
			CCC Pro					624
			CAA Gln 215					672
			ACA Thr					720
			CAG Gln					768
			GTG Val					816
 GGC Gly								822

(2) INFORMATION FOR SEQ ID NO:15:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 274 amino acids
 - (B) TYPE: amino acid
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys 1 5 10

Lys Tyr Pro Ser Cys Gly Gly Leu Leu Gln Ser Pro Ile Asp Leu His 20 25 30

Ser Asp Ile Leu Gln Tyr Asp Ala Ser Leu Thr Pro Leu Glu Phe Gln 35 40

Gly Tyr Asn Leu Ser Ala Asn Lys Gln Phe Leu Leu Thr Asn Asn Gly 50 60

His Ser Val Lys Leu Asn Leu Pro Ser Asp Met His Ile Gln Gly Leu 65 70 75 80

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Gln Ser Arg Tyr Ser Ala Thr Gln Leu His Leu His Trp Gly Asn Pro 85 90 95

Asn Asp Pro His Gly Ser Glu His Thr Val S r Gly Gln His Phe Ala 100 105 110

Ala Glu Leu His Ile Val His Tyr Asn Ser Asp Leu Tyr Pro Asp Ala 115 120 125

Ser Thr Ala Ser Asn Lys Ser Glu Gly Leu Ala Val Leu Ala Val Leu 130 135

Ile Glu Met Gly Ser Phe Asn Pro Ser Tyr Asp Lys Ile Phe Ser His145150155160

Leu Gln His Val Lys Tyr Lys Gly Gln Glu Ala Phe Val Pro Gly Phe 165 170 175

Asn Ile Glu Glu Leu Leu Pro Glu Arg Thr Ala Glu Tyr Tyr Arg Tyr 180 185 190

Arg Gly Ser Leu Thr Thr Pro Pro Cys Asn Pro Thr Val Leu Trp Thr 195 200 205

Val Phe Arg Asn Pro Val Gln Ile Ser Gln Glu Gln Leu Leu Ala Leu 210 215 220

Glu Thr Ala Leu Tyr Cys Thr His Met Asp Asp Pro Ser Pro Arg Glu 225 230 235 240

Met Ile Asn Asn Phe Arg Gln Val Gln Lys Phe Asp Glu Arg Leu Val 245 250 255

Tyr Thr Ser Phe Ser Gln Val Gln Val Cys Thr Ala Ala Gly Leu Ser 260 265 270

Leu Gly

(2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear
- (11) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

CTTTTTTGAT ACCCTTCCTT CTGAA

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(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 986 base pairs

(B) TYPE: nucleic acid (C) STRANDEDNESS: both

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 1..975

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

			GGT Gly						48
			GGC Gly				-		96
			GAC Asp						144
			AAC Asn 55	 	 	 			192
			CTG Leu						240
			ACG Thr						288
			GAG Glu		 	 		-	336
			CAT His						384
			TCA Ser 135						432
			AAT Asn						480

	GTA Val					Ala			TTC Phe	52	ε
	GAG Glu 180									57	ε
	CTG Leu								ACA Thr	62	4
	AAC Asn									672	2
	CTG Leu									720	3
	AAC Asn									766	3
	TTC Phe 260									816	5
	ATC Ile									864	1
	GTG Val									912	2
	AAC Asn									960)
	CAC His	TGAG	GTCC	CC G	;					986	5

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 325 amino acids (B) TYPE: amino acid

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Ser Lys Trp Thr Tyr Phe Gly Pro Asp Gly Glu Asn Ser Trp Ser Lys 1 5 10

Lys	Tyr	Pro	Ser 20	Cys	G1 y	Gl y	Leu	Leu 25	Gln	Ser	Pro	lle	Asp 30	Leu	His
Ser	As p	11e 35	Leu	Gln	Tyr	Asp	Ala 40	Ser	Leu	Thr	Pro	Leu 45	Gl u	Phe	Gln
Gly	Tyr 50	Asn	Leu	Ser	Ala	Asn 55	Lys	Gln	Phe	Leu	Leu 60	Thr	Asn	As n	Gly
His 65	Ser	Val	Lys	Leu	As n 70	Leu	Pro	Ser	Asp	Met 75	His	Ile	Gln	Gly	Leu 80
Gln	Ser	Arg	Tyr	Ser 85	Ala	Thr	Gln	Leu	His 90	Leu	His	Trp	Gly	As n 95	Pro
			100		Ser			105					110		
Ala	Glu	Leu 115	His	Ile	Val	His	Tyr 120	As n	Ser	Asp	Leu	туr 125	Pro	Asp	Ala
Ser	Thr 130	Ala	Ser	Asn	Lys	Ser 135	Glu	G1 y	Leu	Ala	Val 140	Leu	Ala	Val	Leu
11e 145	Glu	Met	Gly	ser	Phe 150	As n	Pro	Ser	Tyr	Asp 155	Lys	Ile	Phe	Ser	His 160
Leu	Gln	His	Val	Lys 165	Tyr	Lys	Gly	Gln	Glu 170	Ala	Phe	Val	Pro	Gly 175	Phe
			180		Leu			185					190		
Arg	Gly	Ser 195	Leu	Thr	Thr	Pro	Pro 200	Суѕ	Asn	Pro	Thr	Val 205	Leu	Trp	Thr
	210				Val	215					220				
225					Cys 230					235					240
				245	Arg				250					255	
_			260		Gln			265					270		
Leu	Gly		Ile		Ser	Leu	Ala 280		Ala	Gly	Ile	Leu 285	Gly	Ile	Суѕ
Ile	Val 290		Val	Val	Ser	11e 295	-	Leu	Phe	Arg	Arg 300		Gly	Ile	Lys

Lys Gly Asp Asn Lys Gly Val Ile Tyr Lys Pro Ala Thr Lys Met Glu 305 310 315

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Thr	Glu Ala His Ala 325	
(2)	INFORMATION FOR SEQ ID NO:19:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 21 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
ACA'	TTGAAGA GCTGCTTCCG G	21
(2)	INFORMATION FOR SEQ ID NO:20:	
	(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 19 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
AAT	TTGCACG GGGTTTCGG	19
(2)	INFORMATION FOR SEQ ID NO:21:	

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1363 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

CTGACACCAC TCAGACCGTG TGTGATCTGG CTCAACCAGT TCTGCGATCC CACCCAGGAA

CAGAAGACTG CAAGAAAACG TTACTTCAAC CCCCCTGTGA TCCCATCTGC AACCTGACCA

ATCAGCACTC CCCAAGTCCC AAGCCCCTAT CTGCCAAATT ATCTTTAAAA ACTCCCCAGA

GGCAGGGTGC AGTGGTTCAA CGCCTGTAAT CCCAGCACTT TAGGTGGATC ACGAGATCAA

GAGATCAAGA CCAGCCTGGC CAACATGGTG AAACCCCGTC TTCTTACTAA AAATACAAAA

ATTAGCTGGG TGTGGCGGCG CGTGCCTGTA ATCCCAGCTA CCCAGGAGGC TGAGGCAGGA

60

120

180

240

300

360

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GAATCGCTTG	AACCCGTGAG	GCAGAGGTTG	CAGTGAGCCA	AGACCATGCC	ACTGCATTTC	420
AGCCTGGGCG	ACAGAGGGGA	ACTCCGTCTG	AACAAACAAA	CAAACAAACA	ACTCCCGGAA	480
TGCTTGGGGA	GACTGATTTG	AGTACTGGAA	TCCCAGTACT	TTAGGAGGCC	AAGGTAGGTG	540
GATCATTTGA	GGTCAGGAGT	TCCAGACCAG	CCTGGCCAAC	ATGGTGAAAC	CCCGTCTCTA	600
CTAAAATTAG	AAAAATTAGC	CGGGTGTGGT	GGTGGGCGCC	TGTAATCCCA	GCACTTTGGG	660
AAGCCAAGGC	AGGTGAATTA	TCTGAGGTCG	GGAGTTTAAG	GCCAGCCTTA	AACTGGCGAA	720
ACCCCGCCTC	TACTAAAAAT	ACAAAAATTA	TCTGGGCATG	GTGGCATGTG	CCTGTAATCC	780
CAGCTACTCG	GGAGGCTGAG	GCAGGAGAAT	CGCTTGAACC	CGGGAGGCGG	AGGTTGCAGT	840
GAGCCGAGAT	CACGCTATTG	CACTCCGGCC	TGGGCAACAG	AGCGAGACTC	CGTCTCAAAC	900
AAACAAACAA	AGGAACGAAA	ACTCCGGTCT	CCGGCACGGC	AAGCTCTGCG	TGAATTACTT	960
TCTCCATTGC	AACTCCCCTG	TCTTGATAAA	TGGGCTCTGT	CTAAGCAGCG	GGCAAGGTGA	1020
ACTCGTTGGG	CTGTTACAGG	ACCAGTGACA	GACCAAGGCA	TGCCACTGAA	GGAATCCCTA	1080
GACGCACCCT	TCTGGATGTG	AGGCAGGCGG	ATCTCACCCC	ACGCCTGCCA	GCAGCTCCTC	1140
GGAGAACTGT	GTTCCTGGGT	CAGCCCTGGC	CCAGAGGAGC	GCCGGGGACC	CGCAGAGTGC	1200
TGCTGAAGTC	AAGGCTACAA	CTCACCTAGG	ATCTGGGGCG	CCAGCCTCCG	GTGGGCAGGG	1260
CGTTCTCCTC	CCCCACCCCC	TCCCCGCACG	ATGACATCAA	GTGTTTGGCG	TTGAGTTGCT	1320
CCATAAAAGC	TGCCCGGGGA	AGCCAGGAGA	GCGAAGGGCG	GAC		1363

(2) INFORMATION FOR SEQ ID NO:22:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs (B) TYPE: nucleic acid

 - (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

GTCCACTTGG ATCCGTTCAC TGG

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WE CLAIM:

1. A substantially purified nucleic acid encoding the amino acid sequence of HCAVIII depicted in SEQ ID ${\tt NO:2.}$

2. The nucleic acid of Claim 1 wherein said nucleic acid is mRNA.

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- 3. A cDNA encoding the amino acid sequence of HCAVIII or a portion thereof.
- 4. The cDNA of Claim 3 wherein the amino acid sequence is encoded by the coding region of the nucleotide sequence depicted in SEQ ID NO:1.
- 5. The cDNA of Claim 3 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:2.
- 6. The cDNA of Claim 3 wherein the amino acid sequence is encoded by the coding region of the nucleotide sequence depicted in SEQ ID NO:3.
- 7. The cDNA of Claim 3 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:4.
- 8. The cDNA of Claim 3 wherein the amino acid sequence is encoded by the nucleotide sequence depicted in SEQ ID NO:12.
- 9. The cDNA of Claim 3 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:13.
- 10. The cDNA of Claim 3 wherein the amino acid sequence is encoded by the nucleotide sequence depicted in SEQ ID NO:14.
- 11. The cDNA of Claim 3 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:15.

- 12. The cDNA of Claim 3 comprising the nucleotide sequence depicted in SEQ ID NO:5.
- 13. The cDNA of Claim 3 comprising the nucleotide sequences depicted in SEQ ID NO:5 and SEQ ID NO:7.

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- 14. A cDNA encoding the amino acid sequence of HCAVIII wherein the phosphorylation region has been mutated.
- 15. The cDNA of Claim 14 wherein the amino acid sequence is encoded by the nucleic acid sequence depicted in SEQ ID NO:17.
- 16. The cDNA of Claim 14 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:18.

- 17. A protein comprising the amino acid sequence of HCAVIII or a portion thereof.
- 18. The protein of Claim 17 wherein the amino acid sequence is encoded by the coding region of the nucleic acid sequence depicted in SEQ ID NO:1.
- 19. The protein of Claim 17 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:2.
- 20. The protein of Claim 17 wherein the amino acid sequence is encoded by the coding region of the nucleic acid sequence depicted in SEQ ID NO:3.
- 21. The protein of Claim 17 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:4.
- 22. The protein of Claim 17 wherein the amino acid sequence is encoded by the coding region of the nucleic acid sequence depicted in SEQ ID NO:12.
- 23. The protein of Claim 17 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:13.
- 24. The protein of Claim 17 wherein the amino acid sequence is encoded by the coding region of the nucleic acid sequence depicted in SEQ ID NO:14.
- 25. The protein of Claim 17 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:15.

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- 26. A protein comprising the amino acid sequence of HCAVIII wherein the phosphorylation region has been mutated.
- 27. The protein of claim 26 wherein the amino acid sequence is encoded by the nucleic acid sequence depicted in SEQ ID NO:17.
- 28. The protein of Claim 26 wherein the amino acid sequence comprises the sequence depicted in SEQ ID NO:18.

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29. A recombinant DNA clone comprising a cDNA of a HCAVIII transcript isolatable from human A549 cells of about 1.1 kilobases.

- 30. An expression vector comprising the nucleic sequence for HCAVIII or a portion thereof.
- 31. The expression vector of Claim 30 wherein the nucleic acid sequence comprises the coding region of the nucleotide sequence depicted in SEQ ID NO:1.
- 32. The expression vector of Claim 30 wherein the nucleic acid sequence comprises the coding region of the nucleotide sequence depicted in SEQ ID NO:3
- 33. The expression vector of Claim 30 wherein the nucleic acid sequence comprises the coding region of the nucleotide sequence depicted in SEQ ID NO:12.
- 34. The expression vector of Claim 30 wherein the nucleic acid sequence comprises the coding region of the nucleotide sequence depicted in SEQ ID NO:14.
- 35. The expression vector of Claim 30 wherein the nucleic acid sequence comprises the nucleotide sequence depicted in SEQ ID NO:17.

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- 36. A method of detecting cancerous and precancerous lung tissue comprising:
 - (a) preparing a section of biopsy tissue;
- (b) probing said tissue with a labeled probe complementary to the cDNA of SEQ ID NO:1;
- (c) removing said probe which has not hybridized to the tissue; and
 - (d) detecting the presence of the hybridized probe.

37. A method for detecting lung cancer antigen specific for non-small cell carcinoma in a human cell specimen comprising:

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- a) labeling a DNA probe comprising the genomic clone of HCAVIII;
- b) reacting the labeled DNA probe with a human test cell specimen and a normal human cell specimen under conditions suitable for hybridization of the labeled probe to any HCAVIII mRNA which may be present in the test and normal cell specimen;
- c) removing unreacted components from the test and said normal cell specimens;
- d) detecting the hybridized probe bound to the test and normal cell specimens;
- e) quantifying and comparing the amount of hybridized probe bound to the test and normal cell specimens.
 - 38. The method of claim 37 further comprising:
- a) labeling a DNA probe comprising the genomic clone of HCAVIII with a substrate which can bind to a detecting substance to form a labeled DNA probe;
- b) reacting the labeled DNA probe with a human test cell specimen and a normal human cell specimen under conditions suitable for hybridization of the labeled probe to any HCAVIII mRNA which may be present in the test and normal cell specimens;
- c) removing unreacted components from the test and normal cell specimens;
- d) reacting the test and normal cell specimens with a detecting substance which is capable of fluorescing;
- e) comparing the fluorescence of the test and normal cell specimens.

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39. A method for screening human specimens for HCAVIII protein, comprising:

- a) mixing a human test specimen with a first amount of an antibody specific for the HCAVIII protein in a first reaction well;
- b) mixing a control lung cancer antigen comprising at least a portion of the HCAVIII protein with a second amount of said antibody specific for the HCAVIII protein in a second reaction well; and

c) detecting whether said test specimen binds to said antibody as compared to said control lung cancer antigen.

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40. A method for testing a human cell sample for lung cancer comprising assaying a cell homogenate for carbonic anhydrase activity.

- 41. An antibody made by immunizing animals with a lung cancer antigen associated with non-small cell lung cancer cells.
- 42. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:2.
- 43. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:4.
- 44. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:13.
- 45. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:15.
- 46. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:18.

- 47. A therapeutic composition for the treatment of non-small cell lung cancer comprising an antibody to HCAVIII protein bound to a substance which affects the ability of said cancer to replicate.
- 48. The method of claim 47 wherein said substance is a cancer drug.
- 49. The method of claim 48 wherein said substance is a radioisotope.
- 50. The method of claim 49 wherein said substance affects gene expression of a gene encoding HCAVIII.

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51. A substantially purified nucleic acid comprising the nucleotide sequence depicted in SEQ ID NO:7.

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52. A cDNA comprising the nucleotide sequence depicted in SEQ ID NO:7.

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53. A substantially purified nucleic acid comprising the nucleotide sequence depicted in SEQ ID NO:21.

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AMENDED CLAIMS

[received by the International Bureau on 20 November 1995 (20.11.95); original claim 41 amended; remaining claims unchanged (1 page)]

- 41. An antibody made by immunizing animals with HCAVIII, a lung cancer antigen associated with non-small cell lung cancer cells.
- 42. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:2.
- 43. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:4.
- 44. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:13.
- 45. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:15.
- 46. The antibody of Claim 41 wherein said lung cancer antigen has the amino acid sequence depicted in SEQ ID NO:18.

AMENDED SHEET (ARTICLE 19)

ASPONCIDONCOPE-QNSKLIPIA-NGNNGSPUDIKTSETKHOTSLKPISVS-INPATAKEIINVCHSFHVNFEDNOM -SHHWGIGKHKGPE-HWHKDFPIA-KGENGSPVDIDTHTAKTDFSLRPLSVS-IDGATSLRILNFGHAFNVEFDOSQD -AKEWGIASHKGPD-HWHELFPNA-KGENGSPVELHTKDIRHDPSLQPMSVS-TDGGSAKTILNFGKTCKVVFDDTTD ARSHWCTEVQAESS-NTPCLVPVKMCGNCQKDRQSPINIVTHKAKVDKKLGRPFPSGIDKKQTWTYQNMGHSVHHLLZNK QHVSDWTTSEGALDEAHWPQHYPAC-GOQRQSPINIVTHKAKVDKKLGRPFPSGIDKKQTWTYQNMGHSVHHLLZNK GHKGWGTQ-DDGPSHWHKLYPIA-QOBRQSPINIQMRDSVTDFQLKLSS-TRAKSLSITMCHSVQVDFHDSDDCANQTSNWTLMP-LATVPVSVP-GOT	RSYLKCOPPSDSTRLPOTEREG-STRENGSERTYDCVKTSABLEVAR-WRSAKTSSLAENASKADCLAVICYLAW-KVG-EA KAVLKCOPLDCTTRLIGGTBERGSLDCQGSERTYDKNKTARLHIVR-WNT-KYGDFGKAVQQPDCLAVLGIPLKVG-SA RSHLRCOPLDCTTRLIGGTLANGSSDDHGSERTYDGVKTAABLALVR-WNP-KYHTIKKALKQRDCIAVIGIFLKIG-HE ASISCOG-LDAPTQAKQLALANSDLPTKGSERSLDCEHPANEHRIVHENREKKGTSRWYKKAQDPEDEIAVLAFLVEAGTQV HTVA-DGIVTIAQMEFHWCASSEISGSERTYDGIRBYIEIHVR-YNS-KYKTIDIAQDAPGLAVLAFVYEVKKT-PE RTVYTCGPLEGAFRLAGFEFHWGKUHDVGSERTYDGIRBYIEIHVR-WNSVKYGHTKEAVYGENCLAVIGVFLETG-DE ASGISCOPLEMHTRLAGFEFHWGKUHDVGSERTYDGRAFAELAIVR-WNSVKYQHTKEAVYGENCLAVIGVFLKIG-AH	<pre>MPFLQKVLDALQAIKTKGKRAPPHYLDFSTLL*PSSLDYMTYPGSL#HPPL/ESSYTWIICKESISVSSDLAQP-RSLLSHV KPGLQKVVDVLOSIKTKGKSADPHYLDPRGLL*PSLDYMTYPGSL#TPPLLECYTWIVLACPISVSSDQVLKP-RKLMFHG NGEYQIPLOALDKIKTKGKRAPPHYLDPRGLL*PACRDYMTYGGSFTFFPCKRCIVWLLACPHYTYSSDQMAKL-RSLLSSA NEGPQPLVRALGHIPKPEMSTTMAESSLLDLL*REEKLAHIFKTIGSL#TTPPCKKVVTVFREPIQLHXPSILASPANTANGLILASPANTANGLYREPIQLHXPQILA-YY NTYTSHYISHYISHYLKYQPTALTLAGLDYQDHL*PRKLQNTYTYTHGSLATPPCKENVVTVFREPIQLHXPQILAFRGILAHR HPSHMRLTDALYMVRFKGTKAQPSCYNPKCLL*PSRHYMTYGGLATPPLESSYTWIQKEPYEVAPSQLAP-RTLL*PS NQTLQRLYDILPEIKHKDARANNPPDPSTLL*PTC*DYMTYRAGLATPPCKTFFYTMFYFUNTPYTKHPYGISQDLALL*PSA</pre> NPSYDKIPSHLQHVKYKGQEAFVPGTNIESLL*PRTTALTYRYRAGLATPPCKTFYLMFYPTMPYQISQDLALL*PSA	Ecdmavphynin-rptyplichtyrref Ecepeelavdaw-rpaplicholkabp Ecepeelavdaw-rppplichtropabp Ecepeelavsiw-rppplichtropabp Eceppelasiw-rppplichtrope-buppchicsepptlichteelldteralm Ktihndtropplichtrope-buppchicseppa Edder-huvmitrpppplichtropplichtropera Loeer-huvmitrploplichtropplichtropplichterfis Hddpspreminfropplichtropplichtropplichtropplichtelisialiaicitetevyvvrinlyrresikkedancytypa
HCAI HCAII HCAII HCAII HCAVI HCAVII HCAVII	HCAII HCAIII HCAIII HCAVI HCAVI HCAVII	HCAI HCAII HCAII HCAI HCAVI HCAVI HCAVII	HCAI HCAII HCAII HCAVI HCAVI HCAVI HCAVI

HCAVIII TRMETEARA*

In. ational application No.
PCT/US95/09145

	A. CLASSIFICATION OF SUBJECT MATTER				
	:Picase See Extra Sheet. :Picase See Extra Sheet.				
According to	o International Patent Classification (IPC) or to both n	ational classification and IPC			
	DS SEARCHED				
Minimum d	ocumentation searched (classification system followed	by classification symbols)			
•	536/22.1, 23.1; 530/350, 387.1; 435/4, 6, 7.1, 7.2;				
Documentat	ion scarched other than minimum documentation to the	extent that such documents are included	in the fields searched		
Electronic d	ata base consulted during the international search (nam	ne of data base and, where practicable	search terms used)		
	ee Extra Sheet.				
C. DOC	UMENTS CONSIDERED TO BE RELEVANT				
Category*	Citation of document, with indication, where app	propriate, of the relevant passages	Relevant to claim No.		
A	American Journal of Pathology, Volume 142, Number 1, 36-38 issued January 1993, Kim et al, "Interphase cytogenetics in paraffin sections of lung tumors by non-isotopic in situ				
	hybridization", pages 307-317.				
A	Cancer (supplement), Volume 69, Number 6, issued 15 March 1992, Gray et al, "Molecular Cytogenetics in Human Cancer Diagnosis", pages 1536-1542.				
A	Cancer Research (Supplement), Volume 52, issue 01 May 1992, Birrer et al, "Application of molecular genetics to the early diagnosis and screening of lung cancer", pages 2658s-2664s.				
X Further documents are listed in the continuation of Box C. See patent family annex.					
Special categorise of cited documents: The start document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the premium.					
to to	be of particular relevance whier document published on or after the international filing date	"X" document of particular relevance; it considered novel or cannot be consid	he claimed invention cannot be		
له ۱۰۲۰	comment which may throw doubts on priority claim(s) or which is had to establish the publication date of another citation or other	when the document is taken alone			
cand to establish the publication almost housest consistence of water appeals reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means. "O" decument referring to an oral disclosure, use, exhibition or other means.					
·p-	"P" document published prior to the international filing date but later than "g," document member of the same patent family the priority date claimed				
Date of the actual completion of the international search Date of mailing of the international search report					
12 OCTOBER 1995 250CT1995					
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Authorized officer Dubach Trucks 107 ETHAN WHISENANT					
	on, D.C. 20231 No. (703) 305-3230	Telephone No. (703) 308-1096			
· evenue					

Form PCT/ISA/210 (second sheet)(July 1992)*

Inc...ational application No.
PCT/US95/09145

C (Continue	tion). DOCUMENTS CONSIDERED TO BE RELEVANT	
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
Y	US,A, 5,134,075 (HELLSTROM ET AL) 28 July 1992, especially column 4 lines 34-64.	39, 41, 47-50
Y	US,A, 4,816,402 (ROSEN ET AL) 28 March 1989, see entire document.	39, 41
A	Gastroenterology, Volume 105, Number 3, issued 1993, Mori et al, "The significance of carbonic anhydrase expression in human colorectal cancer", pages 820-826, see abstract.	40
A	DNA and Cell Biology, Volume 11, Number 7, issued September 1992, Skonier et al, "cDNA cloning and sequence analysis of Bigha, a novel gene induced in a human adenocarcinoma cell line after treatment with transforming growth factor-beta", pages 511-522, see entire document.	1-35, 42-46, 51- 53
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Box 1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)		
This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:		
1. Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:		
2. Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:		
Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a)		
Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)		
This International Searching Authority found multiple inventions in this international application, as follows:		
Please See Extra Sheet.		
1. As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.		
2. As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.		
As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:		
· · · · · · · · · · · · · · · · · · ·		
4. No required additional search fees were timely paid by the applicant. Consequently, this international scare restricted to the invention first mentioned in the claims; it is covered by claims Nos.:		
Remark on Protest The additional search fees were accompanied by the applicant's protest. No protest accompanied the payment of additional search fees.		

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Intomational application No. PCT/US95/09145

A.	CLASSIFICATION	OF	SUBJECT	MA	TTER
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IPC (6):

CO7H 19/00, 21/00, 21/02, 21/04; C07K 1/00, 14/00, 17/00, 16/00; C12Q 1/00, 1/68; G01N 33/53, 33/567; A01N 43/04; A61K 31/70

A. CLASSIFICATION OF SUBJECT MATTER:

US CL :

536/22.1, 23.1; 530/350, 387.1; 435/4, 6, 7.1, 7.2; 514/44, 424/85.8

B. FIELDS SEARCHED

Electronic data bases consulted (Name of data base and where practicable terms used):

APS, BIOSIS, CAPLUS, CANCERLIT

search terms: A549, HCAVIII, Human Cancer Antigen VIII, Cell surface antigen, Cell surface marker, Non-small cell lung cancer, Carbonic Anhydrase

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single invention concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees must be paid.

Group I, claim(s)1-39 and 41-53, drawn to nucleic acids encoding the amino acid sequence of HCVIII, the protein expressed thereof, antibodies to the proteins and methods using one of the above.

Group II, claim(s) 40, drawn to a method for testing a human cell sample for lung cancer by assaying a cell homogenate for carbonic anhydrase activity.

Pursuant to 37 CFR § 1.475(d) the additional method(s) beyond the one first method of use are considered to lack unity and are properly separated.

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